

# Lead determination in canned food by square-wave adsorptive cathodic stripping voltammetry

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**ABSTRACT:** A sensitive and rapid method to analyse Pb(II) in canned fish samples was developed using square-wave adsorptive cathodic stripping voltammetry based on the adsorptive accumulation of 8-hydroxyquinoline complexes of Pb(II) onto a hanging mercury drop electrode, followed by reduction of adsorbed species by voltammetric scan with square wave pulse modulation. The optimum conditions were found to be 0.1 M  $\text{CH}_3\text{COONH}_4$  as a supporting electrolyte, pH 7.5, 8-hydroxyquinoline concentration of 15  $\mu\text{M}$ , accumulation potential  $-0.70$  V (versus Ag/AgCl), accumulation time 120 s, scan rate 0.3 V/s, and pulse amplitude 20 mV. Under the optimum conditions a linear calibration graph was obtained in the concentration range of 0.5–90.0  $\mu\text{g/l}$  with correlation coefficient 0.9973, a limit of detection of 0.108  $\mu\text{g/l}$ , and a limit of quantification of 0.360  $\mu\text{g/l}$ . The recovery values were obtained in the range 93.7–95.1%. The relative standard deviations ( $n = 10$ ) at lead concentration 5.0  $\mu\text{g/l}$  was 2%. The method was successfully applied to the determination of lead content in canned fish samples. The concentration of Pb(II) in canned fish samples (wet weight) was found to be in the range of 0.121–0.285  $\mu\text{g/g}$ , which is lower than the limit (1.00  $\mu\text{g/g}$ ) issued by the Ministry of Public Health of Thailand.

**KEYWORDS:** toxic elements, canned fish, electroanalytical techniques

## INTRODUCTION

There is an increasing concern about the quality of food in several parts of the world in terms of contamination. Toxic elements in food have prompted studies on their toxicological effects and their determinations in food<sup>1</sup> especially lead, which is able to contaminate food during harvesting, processing, and packaging<sup>2</sup>.

Fish is widely consumed in many parts of the world by humans because it has high protein content, low saturated fat, and high omega fatty acids known to promote good health. Canned fish in particular is extensively consumed in the developed world because it is convenient and affordable for most working families<sup>3</sup>. Their toxic metal content should be of some concern to human health. Fish may be contaminated by lead during fish growth, transportation, and storage. Contamination of lead may also occur during production handling and canning process<sup>3</sup>. Solder used in can manufacture is an important source of lead contamination in food during canning<sup>4</sup>. Lead is found at high concentration in muscles and organs of fish. It accumulates in the human body by replacing calcium in bones<sup>5</sup>.

Several methods have been investigated to determine lead at trace level including graphite fur-

nace atomic absorption spectrometry<sup>6</sup>, atomic emission spectrometry (AES) especially when coupled with inductively coupled plasma (ICP-AES)<sup>7</sup>, inductively coupled plasma-mass spectrometry<sup>8</sup>, neutron-activation analysis<sup>9</sup>, and X-ray fluorescence spectrometry<sup>10</sup>. However, these techniques have the disadvantages of complicated operation, high cost of maintenance, expensive apparatus, and the requirement of well-controlled experimental conditions.

Electroanalytical techniques especially stripping analysis are well known as excellent procedures to determine trace chemical species with the advantages of low cost, high sensitivity, easy operation, and the ability of element speciation<sup>11</sup>. Stripping analysis is well recognized as one of the most suitable methods for trace metal determination. Its remarkable sensitivity is attributed to a combination of an effective preconcentration step with advanced measurement procedures that generate a high signal-to-background ratio since the metals are preconcentrated into the electrode by factors of 100–1000<sup>12</sup>. Adsorptive cathodic stripping voltammetry (AdCSV) is becoming increasingly popular to determine trace and ultratrace amounts of metal ions<sup>13</sup>. The technique is based upon adsorptive accumulation of the metal ion complex with a suitable ligand by adsorption at the surface of

the electrode scanned in the negative direction<sup>14,15</sup>. The following reduction step, with a negative potential scan, can be used to measure the adsorbed complex. The adsorptive accumulation approach results in an effective preconcentration with short adsorption times (1–5 min) and highly sensitive and selective for trace metal measurements<sup>12</sup>.

The sensitivity in AdCSV is often greater than that in ASV due to the fact that the metal analytes do not dissolve in the mercury but form a complex layer on a mercury film electrode surface. Most AdCSV procedures utilize a hanging mercury drop electrode (HMDE) to measure reducible species, which offers the advantages of self-cleaning, reproducible surface area, and automatic control capability<sup>16</sup>. Because of the great sensitivity enhancement obtained with AdCSV methods, several complexing agents have been studied for the adsorptive collection of Pb(II) complexes on the HMDE including 8-hydroxyquinoline<sup>17</sup>, xylenol orange<sup>18</sup>, calcein blue<sup>19</sup>, morin<sup>20</sup>, and thymolphthalexone<sup>21</sup>.

8-hydroxyquinoline (oxine) is a well known complexing agent for analytical determination of a number of cations of transition metals<sup>22</sup>. Since 8-hydroxyquinoline can be adsorbed on mercury, it is used in a preconcentration step for labile and non-labile complexes in electroanalytical procedures<sup>23</sup>.

An adsorptive cathodic stripping voltammetric technique was developed here to obtain optimized condition for trace measurement of lead based on effective accumulation of lead(II) complex with 8-hydroxyquinoline on a hanging mercury drop electrode.

## MATERIALS AND METHODS

### Apparatus

The voltammetric measurements were performed by Autolab PGSTAT100 combined with the GPES software, using a multi-mode electrode with the HMDE mode as working electrode, an Ag/AgCl/3 M KCl as a reference electrode and a Pt wire as an auxiliary electrode. Solutions were stirred during the purging and deposition steps by a rotating polytetrafluoroethylene rod. The electrode cell was equipped with a nitrogen purge tube to remove oxygen prior to sample analysis. Square wave voltammetry experiments were carried out with pulse amplitude 20 mV and a scan rate of 0.3 V/s.

### Reagents and solutions

All the reagents were of the analytical reagent grade and were used without further purifications. De-ionized water was used throughout the investigation.

Glassware was rinsed with 10% (v/v) HNO<sub>3</sub> for 48 h followed by thoroughly rinsing with de-ionized water. Stock standard solution of 1000 µg/l Pb(II) was prepared from 1000 mg/l Pb(II) standard solution (SCP Science). Stock solution of 0.01 M 8-hydroxyquinoline (Fluka) was prepared by dissolving 1.11 g 8-hydroxyquinoline in 0.2 M HCl and then diluted with de-ionized water in a 50 ml volumetric flask. The supporting electrolyte was 0.1 M ammonium acetate which was adjusted to obtain pH 7.5 by 0.1 M ammonium hydroxide solution.

### Sample preparation and digestion

Canned fish samples (mackerel in tomato sauce) of ten brands were purchased from local supermarkets. After opening the can, the fish and tomato sauce inside was homogenized thoroughly in a food blender. The homogenized sample (1.5 g wet weight) was placed into a beaker and 15 ml of HNO<sub>3</sub>: perchloric acid: H<sub>2</sub>SO<sub>4</sub> mixture (25:25:1 v:v:v) was added. The beaker was covered by watch glass and heated on a hot plate at 150 °C until the solution was clear. The clear solution was allowed to cool, transferred into a 25 ml volumetric flask, and diluted to the mark with deionized distilled water<sup>5</sup>.

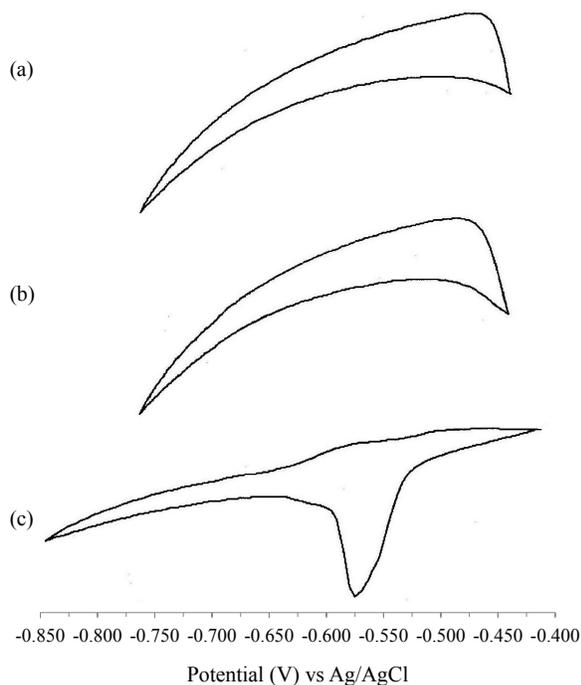
### General voltammetric procedure for sample solution

The sample solution (10 ml) containing 15 µM 8-hydroxyquinoline and 0.1 M CH<sub>3</sub>COONH<sub>4</sub> (pH 7.5) was pipetted into a voltammetric cell. The stirrer was switched on and the solution was purged with nitrogen gas for 1 min. After forming a new HMDE, accumulation proceeded for 120 s at –0.7 V while stirring. At the end of the accumulation time, the stirrer was switched off and the solution was allowed to become quiescent for 10 s. The voltammogram was then recorded by applying a negative potential differential pulse scan.

## RESULTS AND DISCUSSION

### Adsorptive characteristics of the Pb(II)-8-hydroxyquinoline complex

Preliminary experiments were performed to characterize the suitability of 8-hydroxyquinoline for the determination of lead ion using HMDE. Various stripping voltammograms are displayed in Fig. 1. All voltages are reported versus Ag/AgCl. Fig. 1a displays the voltammogram of 0.1 mM 8-hydroxyquinoline solution in 0.01 M ammonium acetate at pH 8.0 after 1 min accumulation at –0.4 V. Fig. 1b shows the voltammogram of a solution containing 1 mg/l

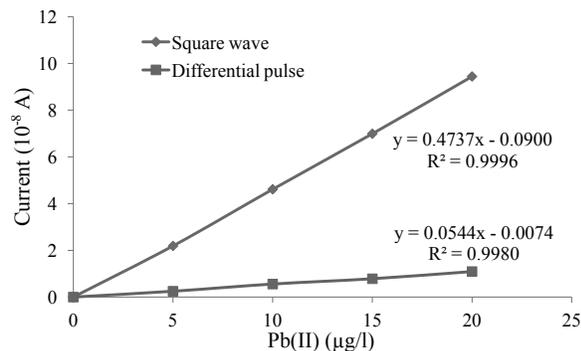


**Fig. 1** Stripping voltammogram of (a) 0.1 mM 8-hydroxyquinoline, (b) 1 mg/l Pb(II), and (c) mixture of 0.1 mM 8-hydroxyquinoline and 1 mg/l Pb(II) in 0.01 M ammonium acetate at pH 8.0 after 1 min accumulation at  $-0.4$  V and scan rate of 50 mV/s.

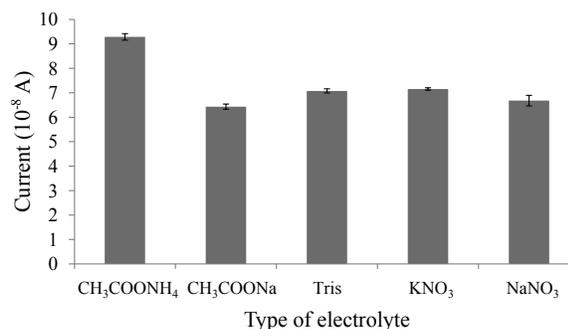
Pb(II) in the absence of 8-hydroxyquinoline ligand under conditions similar to those in Fig. 1a. Fig. 1c shows the voltammogram of a mixture of 0.1 mM 8-hydroxyquinoline and 1 mg/l Pb(II) in 0.01 M ammonium acetate at pH 8.0 after 1 min accumulation at  $-0.4$  V; a reduction peak at  $-0.58$  V was found. It can be concluded that the sensitivity of lead reduction currents is enhanced by the addition of 8-hydroxyquinoline to the solution, indicating that the Pb(II)-8-hydroxyquinoline complex was absorbed on the surface of electrode.

### The comparison between square wave and differential pulse

A number of different waveforms have been used for the stripping step, including linear sweep voltammetry, differential pulse voltammetry (DPV), and square wave voltammetry (SWV). However, SWV and DPV are more commonly used due to their lower detection limits<sup>15</sup>. A comparison of the sensitivities for lead analysis between square wave and differential pulse is shown in Fig. 2. The square wave was found to have greater sensitivity than the differential pulse and was selected for all experiments.



**Fig. 2** The comparison of peak current between square wave and differential pulse of Pb(II) in 0.01 M ammonium acetate containing 10  $\mu$ M 8-hydroxyquinoline at pH 8.0.



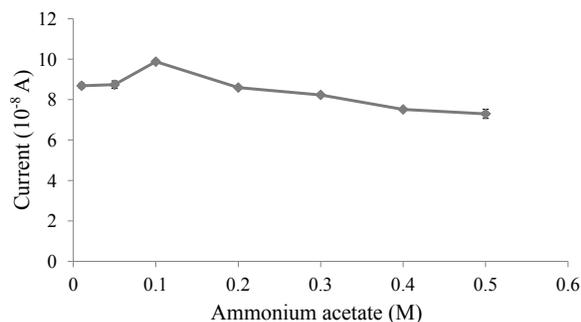
**Fig. 3** Effect of supporting electrolyte on the peak current of 20  $\mu$ g/l Pb(II) in the presence of 10  $\mu$ M 8-hydroxyquinoline at pH 8.0.

### Effect of the supporting electrolyte

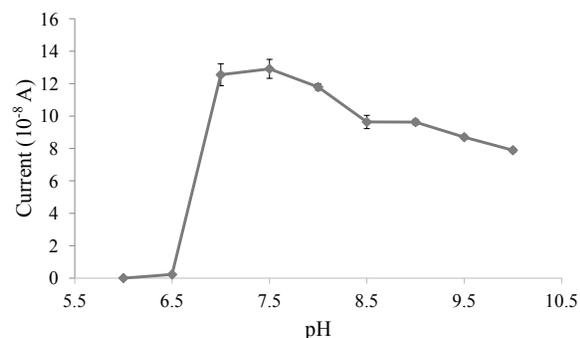
Electrochemical measurements are commonly carried out in a variety of supporting electrolytes to decrease the resistance of the solution, to reduce the effect of migration, and to maintain a constant ionic strength<sup>12</sup>. The effects of different supporting electrolytes including 0.01 M of  $\text{CH}_3\text{COONH}_4$ ,  $\text{CH}_3\text{COONa}$ , Tris,  $\text{KNO}_3$ , and  $\text{NaNO}_3$  are shown in Fig. 3. The highest peak height was achieved in  $\text{CH}_3\text{COONH}_4$  solution. Thus  $\text{CH}_3\text{COONH}_4$  was used as a supporting electrolyte for further experiments.

### Effect of supporting electrolyte concentration

The effect of concentration of the supporting electrolyte on the stripping peak current of Pb(II) was studied by varying the concentration of ammonium acetate within the range of 0.01–0.5 M (Fig. 4). The maximum peak current was observed in 0.1 M ammonium acetate. The increasing of the concentration of ammonium acetate was found to decrease the peak current due to the formation of a weak complex between acetate with Pb(II)<sup>24</sup>. Consequently, an



**Fig. 4** Effect of ammonium acetate concentration on the peak current of 20 µg/l Pb(II) in the presence of 10 µg/l 8-hydroxyquinoline at pH 8.0.



**Fig. 5** Effect of pH on the peak current of 20 µg/l Pb(II) in 0.1 M ammonium acetate containing 10 µM 8-hydroxyquinoline.

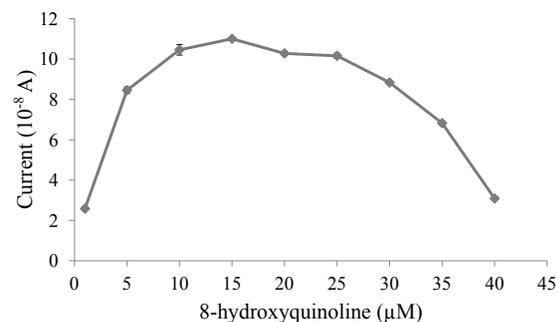
optimum ammonium acetate concentration of 0.1 M was selected for the next experiments.

### Effect of pH

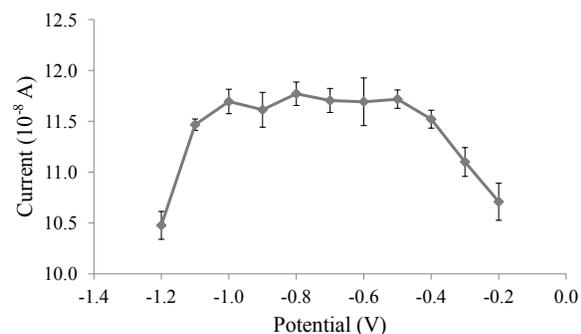
The influence of pH on the stripping peak current of Pb(II) was studied in the pH range of 6.0–10.0 with the results shown in Fig. 5. It indicates that in the pH range of 6.0–7.5, the peak current of the lead complex increases with the increasing of pH to reach the maximum at pH 7.5 and then decreases from pH 8.0–10.0. Thus pH 7.5 was chosen for further studies. At very low pH, the protonation of –NH groups in 8-hydroxyquinoline occurs whereas at high pH the hydrolysis of Pb(II) increasingly affects the formation of Pb(II) and 8-hydroxyquinoline complexes<sup>25</sup>; therefore, at lower or higher pH the complexation of 8-hydroxyquinoline with Pb(II) ions decreases.

### Effect of 8-hydroxyquinoline concentration

The effect of the 8-hydroxyquinoline concentration on the cathodic stripping peak current of 20 µg/l Pb(II) in 0.1 M ammonium acetate at pH 7.5 with an



**Fig. 6** Effect of 8-hydroxyquinoline concentration on the peak current of 20 µg/l Pb(II) in 0.1 M ammonium acetate at pH 7.5.



**Fig. 7** Effect of accumulation potential on the peak current of 20 µg/l Pb(II) in 0.1 M ammonium acetate at pH 7.5.

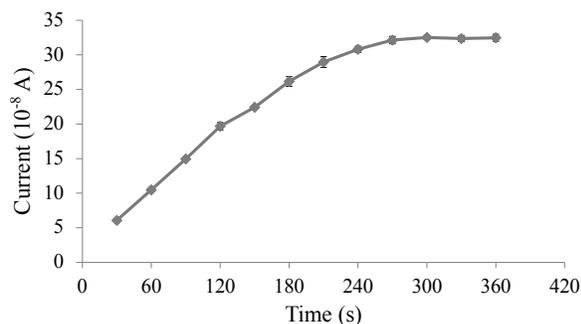
accumulation potential of –1.1 V for 60 s is shown in Fig. 6. The stripping peak current for Pb(II) increased up to 15 µM and then decreased due to the competition of 8-hydroxyquinoline with Pb(II)-8-hydroxyquinoline complexes in adsorption onto the mercury drop electrode<sup>14</sup>. Hence the 8-hydroxyquinoline concentration of 15 µM was selected as an optimum value for further experiments.

### Effect of accumulation potential

The effect of varying accumulation potential on the peak current for Pb(II) determination is shown in Fig. 7. The accumulation potential was varied between –1.2 and –0.2 V. The obtained results revealed that the peak current of Pb(II) was constant between –0.8 V to –0.5 V. Thus an accumulation potential of –0.7 V was selected for lead accumulation because the peak at –0.7 V exhibits constant current trend and provides better sensitivity.

### Effect of accumulation time

The effect of varying accumulation time on the peak current for Pb(II) determination is shown in Fig. 8. It



**Fig. 8** Effect of accumulation potential on the peak current of 20  $\mu\text{g/l}$  Pb(II) in 0.1 M ammonium acetate containing 15  $\mu\text{M}$  8-hydroxyquinoline at pH 7.5.

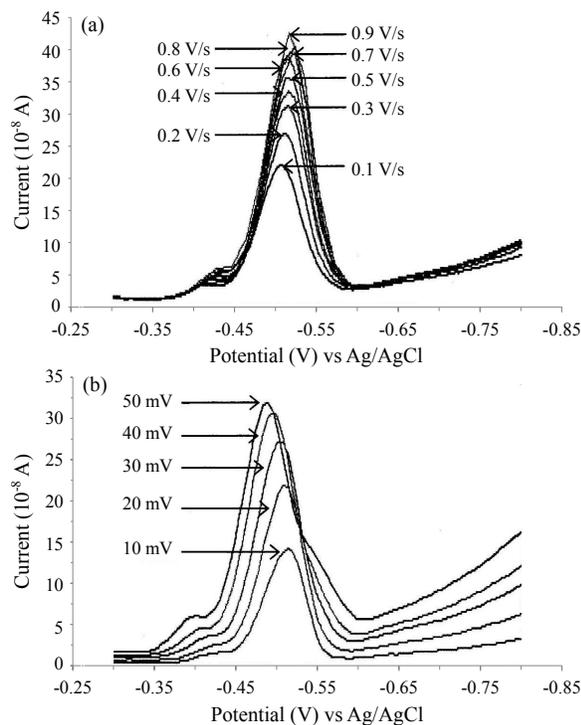
was found that the peak current of Pb(II) increased linearly with the accumulation time, gradually levelling off at periods longer than 270 s which is presumably due to the saturation of the HMDE surface at longer accumulation time<sup>26</sup>. Thus an adsorption time of 120 s was used throughout this work as it combines good sensitivity with relatively short analysis time.

#### Effect of scan rate and pulse amplitude

To improve the sensitivity for the determination of Pb(II), the influences of parameters of square wave voltammetry on the measurement of lead were investigated. The effect of scan rate and pulse amplitude on the peak current in the range of 0.1–0.9 V/s and 10–50 mV is shown in Fig. 9a,b. The peak current for lead increased with increasing scan rate and pulse amplitude; therefore, a scan rate of 0.3 V/s and pulse amplitude 20 mV were selected as the criteria for better sensitivity and peak shape.

#### Analytical performance

At the optimized conditions, the linear calibration graph was obtained in the concentration range of 0.5–90.0  $\mu\text{g/l}$  with the correlation coefficient of 0.9973. The limit of detection (LOD) and the limit of quantification (LOQ) were calculated by carrying out the above procedure with 10 blank samples and calculating  $(3 \times \text{SD})/m$  for LOD and  $(10 \times \text{SD})/m$  for LOQ, where SD is the standard deviation of blank and  $m$  is the slope of calibration graph, which were found to be 0.108  $\mu\text{g/l}$  and 0.360  $\mu\text{g/l}$ , respectively. The accuracy of the determination was tested by spiking the canned fish samples before sample digestion with various concentrations of Pb(II). The percent recovery values were found to be in the range of 93.7–95.1% (Table 1). The analytical precision of the method was estimated from the reproducibility



**Fig. 9** Square wave voltammogram of varied scan rates (a) and pulse amplitudes (b) on the peak current of 20  $\mu\text{g/l}$  Pb(II) in 0.1 M ammonium acetate containing 15  $\mu\text{M}$  8-hydroxyquinoline at pH 7.5.

**Table 1** The percent recovery of Pb(II) at spiked concentration of 10, 20, and 30  $\mu\text{g/l}$  in canned fish.

Sample Type	Pb(II) concentration ( $\mu\text{g/l}$ )	%Recovery
sample	9.6	–
+ 10 $\mu\text{g/l}$ Pb(II)	19.0	94.0
+ 20 $\mu\text{g/l}$ Pb(II)	28.3	93.7
+ 30 $\mu\text{g/l}$ Pb(II)	38.1	95.1

of 10 determinations at three lead concentrations. The relative standard deviations at lead concentrations of 1.0, 5.0, and 10.0  $\mu\text{g/l}$  were 6%, 2%, and 2%, respectively.

#### Interferences

Possible interference by other metals with the adsorptive cathodic stripping voltammetry determination of Pb(II) was investigated by the addition of the interfering ion to a solution containing 20.0  $\mu\text{g/l}$  of Pb(II) and were carried out the measurements under optimized conditions with the results summarized in Table 2. Several ions including  $\text{Fe}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Mn}^{2+}$ , and  $\text{Hg}^{2+}$  (5-fold concentration);  $\text{Sn}^{2+}$  (3-fold concentration);  $\text{Cd}^{2+}$ ,  $\text{Zn}^{2+}$ , and  $\text{Al}^{3+}$  (equal concentration); have

**Table 2** Change in peak current of 20 µg/l Pb(II) in the presence of other ions.

Metal	Concentration (µg/l)	Change in peak current (%)
Pb <sup>2+</sup>	100	-0.3
Mn <sup>2+</sup>	100	7.7
Cr <sup>3+</sup>	100	4.3
Hg <sup>2+</sup>	100	-1.4
Sn <sup>2+</sup>	60	-10.8
Cd <sup>2+</sup>	20	-13.7
Zn <sup>2+</sup>	20	-8.2
Al <sup>3+</sup>	20	-12.9
Cu <sup>2+</sup>	20	-32.9
Ni <sup>2+</sup>	20	-29.5

**Table 3** Pb(II) concentration in canned fish samples by standard addition method.

Sample number	Pb concentration (µg/g)
1	0.160
2	0.285
3	0.208
4	0.180
5	0.259
6	0.151
7	0.140
8	0.265
9	0.121
10	0.277

only negligible effect on the determination of Pb<sup>2+</sup>. Although equal concentrations of Cu<sup>2+</sup> and Ni<sup>2+</sup> interfere significantly by decreasing the Pb<sup>2+</sup> signal, the peak of Pb<sup>2+</sup> is still well separated.

### Applications

The proposed method was applied to the determination of Pb(II) in canned fish samples. The standard addition method was used to eliminate the matrix effect. As shown in Table 3, the concentration of Pb(II) in canned fish samples (wet weight) was found in the range of 0.121–0.285 µg/g, which were lower than standard limited for food contamination (< 1.0 µg/g) issued by the Ministry of Public Health of Thailand.

### Conclusion

The present study demonstrates that square wave adsorptive cathodic stripping voltammetry of lead based on accumulation of Pb(II)-8-hydroxyquinoline complex can be used to determine trace amounts of lead in canned fish samples. This method is simple, sensitive, inexpensive and rapid with the optimum condition of 0.1 M ammonium acetate at pH 7.5 as supporting

electrolyte, 15 µM 8-hydroxyquinoline, accumulation potential -0.70 V, adsorption time 120 s, scan rate 0.3 V/s, and pulse amplitude of 20 mV. Most metal ions have negligible interference effect except Cu<sup>2+</sup> and Ni<sup>2+</sup>. LOD and LOQ are 0.108 µg/l and 0.360 µg/l, respectively, with satisfactory recovery of 93.7–95.1% and reproducibility of 2% for 5.0 µg/l Pb(II).

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