

Toxicity and Accumulation of Lead and Cadmium in the Filamentous Green Alga *Cladophora fracta* (O.F. Müller ex Vahl) Kützing: A Laboratory Study

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ABSTRACT: The toxicity and accumulation of the heavy metals, lead (Pb) and cadmium (Cd) in a common filamentous green alga, *Cladophora fracta*, were studied. *C. fracta* were cultured in a modified Chu No. 10 medium, which was supplemented with 5, 10, 20, 40 or 80 mg/L of Pb or 0.5, 1, 2, 4 or 8 mg/L of Cd, and were separately harvested after 2, 4, 6 and 8 days. The toxicity symptoms of Pb and Cd to *C. fracta* showed damage and reduced number of chloroplasts, disintegrated cell wall and death. There were significant decreases in the relative growth and total chlorophyll content when the exposure time and concentration were increased. The accumulation study showed that there were significant increases of metal levels in algal tissue when the exposure time and concentration were increased. The bioconcentration factor (BCF) of Pb was higher than that of Cd at the same duration, suggesting that the accumulation potential of *C. fracta* for Pb was higher than that for Cd.

KEYWORDS: *Cladophora fracta*, lead, cadmium, toxicity, accumulation.

INTRODUCTION

Water pollution by heavy metals in industrial waste effluents is now a global problem¹. The non-degradability of inorganic pollutants like heavy metals creates a hazard when they are discharged into a water body. The main sources of heavy metal pollution are mining, milling and surface finishing industries, which discharge a variety of toxic metals into the environment². Industrial effluents may be discharged directly into the sea, or into waterways or sewer but whatever the disposal route, these constitute an important source of contamination of the environment³. Many industries discharge the heavy metals lead (Pb) and cadmium (Cd) in their wastewaters⁴. Lead and cadmium are toxic heavy metals and are considered non-essential for living organisms. They are being used in a wide variety of industrial processes in Thailand, for example, the use of Pb in battery, paint and ammunition, and the use of Cd as a coloring agent, a stabilizer and in alloy mixtures. Thus, they were selected as toxicants in the present study.

Traditional technologies for the removal of heavy metals, such as chemical reduction and precipitation or ion exchange, are often ineffective and/or very expensive when used for the removal of heavy metal

ions to very low concentrations. Moreover, these methods are specific to each metal ion. New technologies are required that can reduce heavy metal concentrations to environmentally acceptable levels at affordable costs. Bioremoval offers a potential alternative to existing methods and is defined as the accumulation and concentration of pollutants from aqueous solutions by the use of biological materials⁵. Bioremoval of heavy metals is one of the most promising technologies involved in the removal of toxic metals from industrial waste streams and natural waters. It is a potential alternative to conventional processes for the removal of metals⁶. The major advantages of the bioremoval technology are its effectiveness in reducing the concentration of heavy metal ions to very low levels, and its use of inexpensive biosorption materials and environmentally friendly technologies⁷. Bioremoval is most effective in treatment of waters containing low concentrations of cationic heavy metals. It has been exploited to observe metal contamination in water bodies where the contaminant level is below detection limits.

Algae accumulate heavy metals from their aquatic environment. Using living algae to remove toxic metals from contaminated water could be advantageous, since they are ubiquitous and have colonized almost all parts

of the world. They can be grown easily and have very simple growth requirements. An advantage of using living organisms over dead biomass is that they have fast growth rates and hence produce a regenerating supply of metal – removal material⁸. There is much evidence that algae could accumulate heavy metals in their tissues when grown in polluted waters, including the species *Ulva rigida* (Fe, Mn)⁹, *Padina gymnospora* (Zn)¹⁰, *Gracilaria tenuistipitata* (Cd)¹¹, *Undaria pinnatifida* (Pb)⁴, *Cladophora* sp. (Cd)⁸, and *Cladophora glomerata* (Zn)¹².

Cladophora sp. is a common filamentous green alga in lake shores, rivers and irrigation channels. It is possibly the most ubiquitous freshwater macroalgae worldwide¹³. Metal binding to non-living cells of *Cladophora* sp. is well documented¹⁴⁻¹⁷. However, there are only a few reports on the metal uptake potential of living *Cladophora* sp. and the metal toxicity to this genus^{8,12}. Hence, the purposes of this study were to examine the Pb and Cd accumulation and their toxicities to *Cladophora fracta*.

MATERIALS AND METHODS

Algal Materials

Cladophora fracta (O.F. Müller ex Vahl) Kützing were collected from natural ponds and grown in a modified Chu No. 10 medium¹⁸ in the laboratory under controlled conditions (25 ± 2°C, 45 mmol m⁻²s⁻¹ photon flux intensity, 16h/8h light and dark cycle). The final pH of the solution was 5.0. Two-week-old *C. fracta* was used in the experiments.

Toxicity and Accumulation

The modified Chu No. 10 medium was supplemented with five nominal concentrations of Pb prepared from Pb(NO₃)₂ (5, 10, 20, 40 and 80 mg/L) and Cd prepared from CdCl₂ (0.5, 1, 2, 4 and 8 mg/L). The final pH of the solutions were adjusted to 5.0. One gram fresh weight of algae was inoculated into each flask containing various concentrations of Pb and Cd. Algae cultured in the nutrient medium without heavy metals served as controls. All experiments were performed in triplicate.

Toxicity Symptoms After the exposure of *C. fracta* to Pb and Cd, the algae were harvested at the end of each test duration (2, 4, 6 and 8 days). Toxicity symptoms of treated and control algae were observed under a compound transmission light microscope.

Relative Growth Treated and control algae were gently blotted and weighed after each harvest on day 2, 4, 6 and 8. Relative growth of control and treated algae were calculated as follows:

$$\text{Relative growth} = \frac{\text{Final fresh weight (g)}}{\text{Initial fresh weight (g)}}$$

Total Chlorophyll Content The total chlorophyll content was determined by the absorption spectra of algal extract in a spectrophotometer according to the methods described by Arnon¹⁹ and MacKinney²⁰. The absorbance of the extract was measured at both 663 and 645 nm.

Metal Accumulation The procedures of digestion of algal materials were performed according to Anderson²¹ and Katz and Jennis²². Total accumulations of Pb and Cd in algae were determined using a flame atomic absorption spectrophotometer²³.

Bioconcentration Factor (BCF) The BCF was calculated for quantifying the metal removal potential of the plants. The factor is defined as the ratio of the metal concentration in the dry plant biomass (ppm) to the initial concentration of metal in the feed solution (ppm)²⁴. The BCFs for Pb and Cd of *C. fracta* at different concentrations and exposure times were determined²⁵.

Statistical Analysis

The mean values of relative growth, total chlorophyll content, and metal accumulation were calculated and subjected to analysis of variance (ANOVA) using randomized block design and Least Significant Difference method (LSD) on the SPSS for Windows program after analysis of the homogeneity of variance according to Cochran's test²⁶.

RESULTS

Toxicity Symptoms

The toxicity symptoms observed in both Pb and Cd treatments were rather similar. The symptoms included the damage of chloroplasts, reduction in number of chloroplasts, disintegrated cell wall and cell death. These symptoms were more severe when the metal concentration and exposure time were increased.

Relative Growth

The effects of Pb and Cd on the relative growth of *C. fracta* at different concentrations and exposure times are shown in Figure 1. The relative growth of algae exposed to Pb and Cd at every concentration were significantly decreased (P ≤ 0.05) from those of controls. At high metal concentrations, algal growth was reduced (50%) after 8 days. The lowest relative growth was found in algae treated with Pb at 80 mg/L (0.52; Fig. 1A) and Cd at 8 mg/L (0.47; Fig. 1B) on day 8.

Total Chlorophyll Content

The effects of Pb and Cd on total chlorophyll

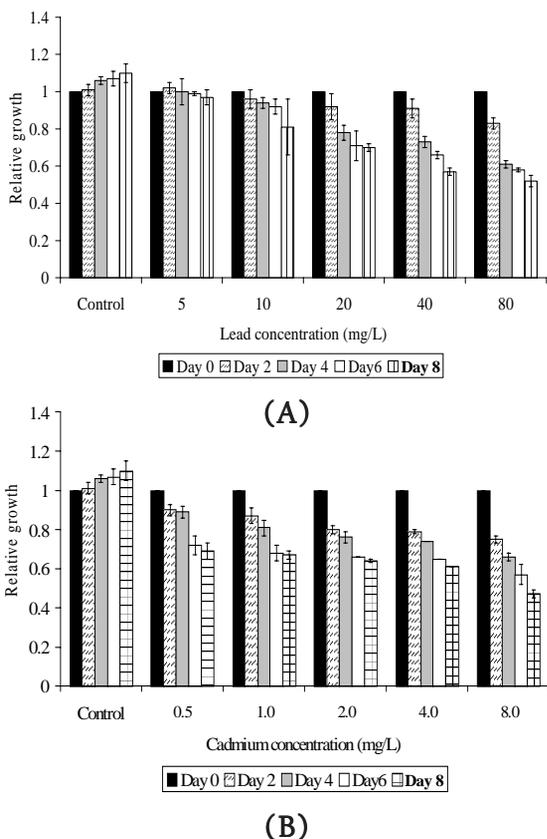


Fig 1. The effects of Pb (A) and Cd (B) on relative growth of *C. fracta* at different concentrations and exposure times.

content of *C. fracta* at different concentrations and exposure times are shown in Figure 2. There were significant decreases ($P \leq 0.05$) of total chlorophyll content when the exposure time and metal concentration were increased. Total chlorophyll contents of *C. fracta* exposed to Pb and Cd at every concentration decreased significantly from those of controls after two days of exposure. The lowest total chlorophyll contents were found in algae exposed to 80 mg/L of Pb (1.0; Fig. 2A) and 8 mg/L of Cd (1.1; Fig. 2B) on day 8.

Metal Accumulation

Pb and Cd accumulations by *C. fracta* at different concentrations and exposure times are shown in Figure 3. There were significant increases ($P \leq 0.05$) of metals in algal tissue when the exposure time and metal concentration were increased. At Pb concentrations of 5, 10, 20, 40 and 80 mg/L, the Pb accumulated in *C. fracta* were 6,170, 11,900, 21,600, 40,000 and 61,400 mg/g dry wt, respectively on day 8 (Fig. 3A). At Cd concentrations of 0.5, 1, 2, 4 and 8 mg/L, the Cd accumulated in *C. fracta* were 603, 1,160, 1,680, 2,630

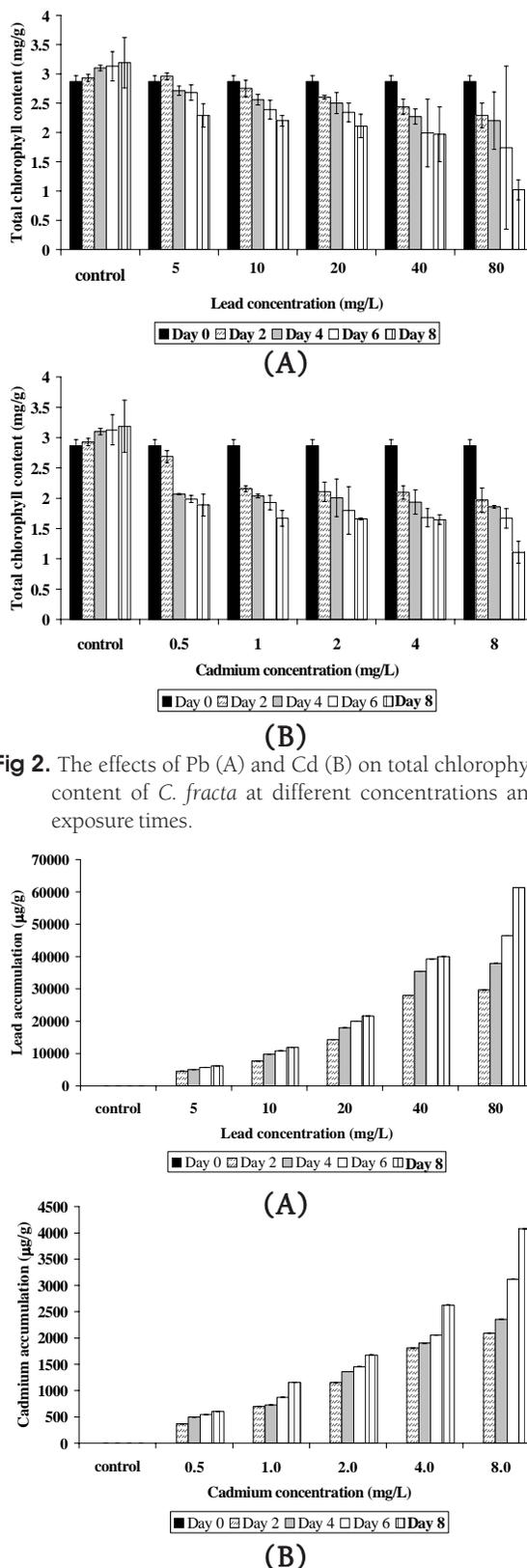


Fig 2. The effects of Pb (A) and Cd (B) on total chlorophyll content of *C. fracta* at different concentrations and exposure times.

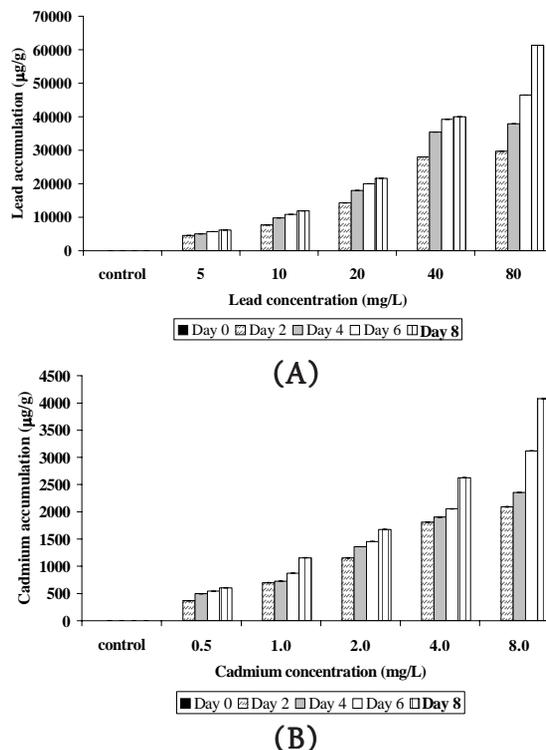


Fig 3. The accumulation of Pb (A) and Cd (B) by *C. fracta* at different concentrations and exposure times.

and 4,080 mg/g dry wt, respectively on day 8 (Fig. 3B). The metals were not detected in the controls.

Bioconcentration Factor

The BCFs for Pb and Cd in *C. fracta* at different concentrations and exposure times are shown in Fig. 4. The BCFs of both metals significantly decreased ($P < 0.05$) when metal concentrations in feed solutions were increased at each exposure time. On day 8, the BCFs of Pb at 5, 10, 20, 40 and 80 mg/L were 1,230, 1,190, 1,080, 1,000 and 767, respectively (Fig. 4A), while those of Cd at 0.5, 1, 2, 4 and 8 mg/L were 1,205, 1,160, 838, 657 and 510, respectively (Fig. 4B).

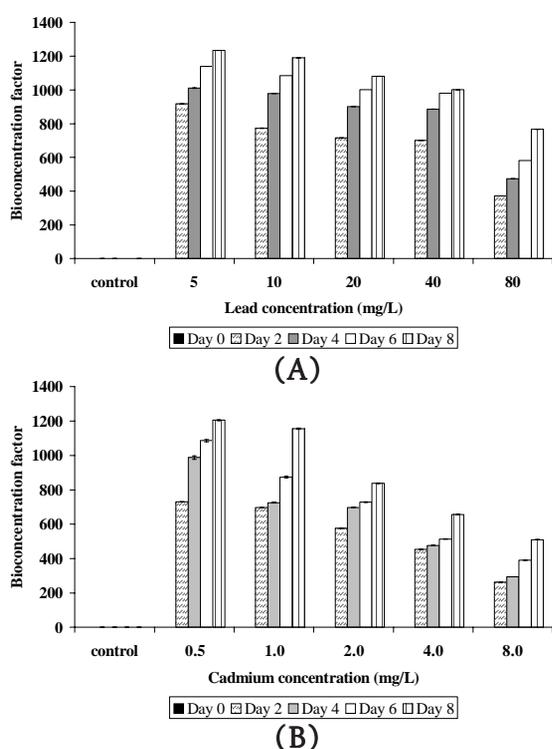


Fig 4. The bioconcentration factor values of Pb (A) and Cd (B) in *C. fracta* at different concentrations and exposure times.

DISCUSSION

In the toxicity symptom study, *C. fracta* exposed to Pb and Cd showed a reduction in number of chloroplasts when compared with those of the control. Cell walls were crumbled and chloroplasts were severely damaged in most cases at high concentrations of metals, but the symptoms were not specific. Similar results were obtained from the toxicity symptom study of metal on *Chlorella*²⁷. *Chlorella* cells were useful in the

characterization of the toxicity of both metals and organic contaminants. The chloroplast is the organelle most affected by metal contamination. In Ni- treated plants (*Brassica oleracea*), the higher the Ni concentration, the smaller the number of chloroplasts in mesophyll cells²⁸. The number and size of chloroplasts decreased, and their internal membranes (especially grana) were reduced and swollen. Uptake and excess of metals by plants and algae can initiate a variety of metabolic reactions, finally leading to global phytotoxic responses, e.g., dwarf growth and chlorosis. They are generally considered to affect membrane permeability and to induce cell decompartmentation. An important harmful effect of metals at the cellular level is the alteration of the plasma membrane permeability, leading to leakage of ions like potassium and other solutes²⁹.

In the present study, the relative growth of *C. fracta* exposed to Pb and Cd decreased significantly when the exposure time and metal concentration were increased. Miranda and Hangovan³⁰ studied the Pb influence on specific growth rate of *Lemna gibba*. They found that high Pb concentrations (200-500 mg/L) in the media significantly inhibited the specific growth rate of *L. gibba* under continuous and discontinuous illumination. This might be due to the fact that Pb induces the activity of the enzyme peroxidase that is involved in the degradation of indoleacetic acid (IAA), the hormone which stimulates plant growth and multiplication.

Several studies have reported on the effects of Cd and algal growth. Fargasova³¹ studied the effects of Cd, Cu, Zn, Pb and Fe on the green alga *Scenedesmus quadricauda* and found that the toxicity for all the observed parameters increased with the concentration of these metals in the cultivation medium. Lasheen et al³² reported that Cd had slight inhibitory effects on algal growth at low concentration (0.05 mg/L), while it severely inhibited algal growth at higher concentrations (>1.0 mg/L). Leborans and Novillo³³ found that Cd caused a decrease of the cellular volume, the growth rate and of the level of photosynthetic pigments.

The total chlorophyll content of *C. fracta* significantly decreased when the exposure time and Pb or Cd concentration were increased. Pb and Cd at high concentrations destroyed chloroplasts of *C. fracta*, as shown in the toxicity symptom study. It is well known that Cd can cause disorganization of chloroplasts leading to a reduction of the photosynthetic pigments³³. Both Cd and Pb were reported to inhibit chlorophyll biosynthesis, leading to the lowered chlorophyll contents³⁴. Sen and Mondal³⁵ reported that the decline in chlorophyll content might be caused by a reduction in the synthesis of chlorophyll, possibly by increasing chlorophyllase activity, by disorderness of chloroplast membrane and by inactivation of electron transport in photosystem I.

C. fracta possess the potential to accumulate metals in their tissues. The results revealed that, under the experimental condition, the accumulations of Pb and Cd by *C. fracta* were increased when the exposure time and metal concentration were increased. Similarly, Costa and Leite³⁶ found that the amount of Cd and Zn accumulated by *Chlorella* was dependent on the external metal concentration, with increasing metal accumulation at increased external metal concentrations. The total Pb sorption by *Stichococcus bacillaris* cells increased with the increasing external Pb concentration and time of exposure³⁷. Maine et al³⁸ also concluded that both initial Cd concentration and time had a statistically significant effect on the sorption of Cd by *Pistia stratiotes*.

In the present study, *C. fracta* accumulated Pb and Cd to the highest concentrations of 61,400 mgPb/g when exposed to 80 mg Pb/L, and 4,090 mgCd/g when exposed to 8 mg Cd/L (Figure 5). Several studies have found high levels of metal accumulation. Water milfoil (*Myriophyllum spicatum*) exposed to 16 mg Pb/L and 16 mg Cd/L could accumulate 36,500 mgPb/g and 2,800 mgCd/g, respectively³⁹. *Chlorella vulgaris* accumulated 5,000 mgCd/g when exposed to 50 mM

Cd⁴⁰. Tokunaga et al⁴¹ found that a water hyacinth exposed to 5 mg/L of Cd attained in an ultimate leaf concentration of 1,010 mgCd/g and a root concentration of 2,230 mgCd/g. In comparison, other algae and aquatic plant species were proven to be poor accumulators of metals. Zayed et al⁴² reported that the highest concentrations of each trace element accumulated in duckweed tissue were 13.3 mgCd/g and 0.63 mgPb/g when treated with 10 mg Cd/L and 10 mg Pb/L, respectively. Bulrush (*Scirpus robutus*) and saltmeadow cordgrass (*Spartina patens*) accumulated 200 and 250 mgCd/g when exposed to 0.5 and 1.0 mg/L Cd, respectively⁴³. In comparison, *C. fracta* can be considered a good accumulator for Pb and Cd.

BCF is a useful parameter to evaluate the potential of plants for accumulating metals and this value was calculated on a dry weight basis²⁴. In this study, the BCF values of *C. fracta* in each group of Pb were significantly higher than those in each group of Cd, indicating that the uptake of Pb was better than that of Cd. There was a gradual decrease in the Pb and Cd uptake potential with an increase in Pb and Cd concentration in feed solutions. The ambient metal concentration in water is the major factor influencing the metal uptake efficiency⁴⁴⁻⁴⁵ and concomitantly the BCF values, as found in the present study. Similar experiments and similar results have also been reported by Zhu et al⁴⁶ who found that BCFs of water hyacinth were very high for Cd, Cu, Cr and Se at low external concentration, and they were decreasing as the external concentration increased. Zayed et al⁴² reported that duckweed bioconcentrated the six elements (Cu, Se, Pb, Cd, Ni, Cr) under study to different levels at low supply concentrations compared with those at high supply concentrations. Studies have also found high levels of Cd accumulation in floated duckweed, *L. gibba* (5,953)⁴⁷ and ivy duckweed, *L. trisulca* (3,594)⁴⁸. Rai et al⁴⁹ reported Cd BCF values ranging from 2,125 to 29,000 for six wetland plant species (coontail, giant duckweed, bacopa, wild rice, channel grass) and green algae (*Hydrodictyon reticulatum* and *Chara corallina*). Zhu et al⁴⁶ found the Cd BCF value of 2,150 for water hyacinth. From the view of phytoremediation, a good accumulator should have the ability to concentrate the elements in its tissue, for example, a BCF of more than 1,000 (100-fold compared on a fresh weight basis)⁴². Based on this criterion, our results showed that *C. fracta* is a good accumulator of Pb and Cd with BCF values of 1,234 and 1,205, respectively.

Requirements for developing a practical bioremoval process for heavy metals include low-cost production of plant biomass, ease of removing the biomass from suspension, high maximum specific adsorption, and the capability to reduce metal concentration to very low residual levels. The green macroalgae *C. fracta* are

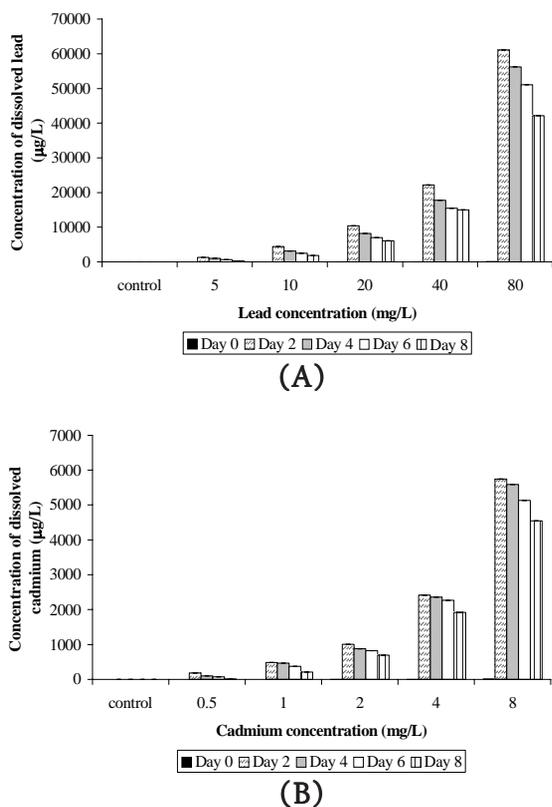


Fig 5. Concentrations of dissolved Pb (A) and Cd (B) remaining in the culture media of *C. fracta* at different concentrations and exposure times.

very easy to harvest and are potentially produced in mass culture. The experimental data presented here indicated that these algae may have promising metal adsorbing characteristics. The fact that *C. fracta* had high BCFs for Pb and Cd at low external concentration is also important for phytoremediation because, to its advantage, the process is more cost-efficient than other conventional techniques in treating large volumes of wastewater with low concentrations of pollutants. However, more work is needed to optimize the design and management of an aquatic plant based system so as to get maximum efficiency in metal removal. The system should have a confined environment such as a constructed wetland or lagoon system. In addition, knowledge of water chemistry, presence of humic acid in the system, harvesting techniques, metal recovery technology, and safe disposal of used plants will have to be worked out before large scale application is adopted.

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REFERENCES

- Jana S (1988) Accumulation of Hg and Cr by three aquatic species and subsequent changes in several physiological and biochemical plant parameters. *Water Air Soil Pollut* **38**, 105-9.
- Figueira MM, Volesky B, Ciminelli VST and Roddick F A (2000) Biosorption of metals in brown seaweed biomass. *Water Res* **34**, 196-204.
- Srivastav RK, Gupta SK, Nigam KDP and Vasudevan P (1994) Treatment of chromium and nickel in wastewater by using aquatic plants. *Water Res* **7**, 1613-38.
- Kim YH, Park JY, Yoo YJ and Kwak JW (1999) Removal of lead using xanthated marine brown alga, *Undaria pinnatifida*. *Process Biochem* **34**, 647-52.
- Sag Y and Kutsal T (1997) The simultaneous biosorption process of lead (II) and nickel (II) on *Rhizopus arrhizus*. *Process Biochem* **32**, 591-7.
- Veglio F, Beolchini F and Gasbarro A (1995) Biosorption of toxic metals: an equilibrium study using free cells of *Arthroabacter* sp. *Process Biochem* **32**, 99-105.
- Yu Q, Matheickal JT, Yin P and Kaewsarn P (1999) Heavy metal uptake capacities of common marine macroalgal biomass. *Water Res* **33**, 1534-37.
- Sobhan R and Sternberg SPK (1999) Cadmium removal using *Cladophora*. *J. Environ Sci Health Part A* **34**, 53-72.
- Favero N, Cattalini F, Berlaggia D and Albergoni V (1996) Metal accumulation in a biological indicator (*Ulva rigida*) from the lagoon of Venice (Italy). *Arch Environ Contam Toxicol* **31**, 9-18.
- Karez CS, Magallhaes VT, Pfrlffer WC and Filho GMA (1994) Trace metal accumulation by algae in Sepatiba Bay, Brazil. *Environ Pollut* **83**, 351-6.
- Hu SX, Tang CH and Wu ML (1996) Cadmium accumulation by several seaweeds. *Sci Total Environ* **187**, 65-71.
- McHardy BM and George JJ (1990) Bioaccumulation and toxicity of zinc in the green alga, *Cladophora glomerata*. *Environ Pollut* **66**, 55-66.
- Planas D (1996) Phosphorus and nitrogen relationships of *Cladophora glomerata* in two lake basins of different trophic status. *Freshwater Biol* **35**, 602-22.
- Nourbakhsh M, Sag Y, Ozer D, Aksu Z, Kutsal T and Caglar A (1994) A comparative study of various biosorbents for removal of chromium (VI) ions from industrial waste waters. *Process Biochem* **29**, 1-5.
- Ozer D, Aksu Z, Kutsal T and Caglar A (1994) Adsorption isotherm of lead (II) and chromium (VI) on *Cladophora crispata*. *Environ Technol* **15**, 439-48.
- Aksu Z, Ozer D, Ekiz HI, Kutsal T and Caglar A (1996) Investigation of biosorption of chromium (VI) on *Cladophora crispata* in two- staged batch reactor. *Environ Technol* **17**, 215-20.
- Aksu Z and Kutsal T (1998) Determination of kinetic parameters in the biosorption of copper (II) on *Cladophora* sp., in a packed bed column reactor. *Process Biochem* **33**, 7-13.
- Bold HC (1980) Growth media recipes. (Online). Available URL: www.bio.utexas.edu/research/utex/media/chu.html.
- Arnon DI (1949) Copper enzyme in isolated chloroplasts polyphenol oxidases in *Beta vulgaris*. *Plant Physiol* **24**, 1-14.
- MacKinney G (1941) Absorption of light by chlorophyll solutions. *J Biol Chem* **144**, 315-23.
- Anderson R, (1991) *Sample Pretreatment and Separation*, pp 1-1632. John Wiley & Sons, Singapore.
- Katz SA and Jennis SW (1983) *Regulatory Compliance Monitoring by Atomic Absorption Spectroscopy*, pp 1-285. Verlac Chemie International Inc, USA.
- APHA, AWWA, WEF (1998) *Standard Methods for the Examination of Water and Wastewater*, 20th ed, Washington DC.
- Raskin I, Kumar PBAN, Dushenkov S and Salt D (1994) Bioremediation of heavy metals by plants. *Curr Opin Biotechnol* **28**, 115-26.
- Jain SK, Vasudevan P and Jha NK (1990) *Azolla pinnata* R. Br. and *Lemna minor* L. for removal of lead and zinc from polluted water. *Water Res* **24**, 177-83.
- Winer BJ (1981) *Statistical Principles in Experimental Design*. International Student Edition, pp 1-907. McGraw-Hill, New York.
- Wong SI, Nakamoto L and Wainwright, JF (1997) Detection of toxic organometallic complexes in wastewaters using algal assays. *Arch Environ Contam Toxicol* **322**, 358-66.
- Molas J (1997) Changes in morphological and anatomical structure of cabbage (*Brassica oleracea* L.) outer leaves and in ultrastructure of their chloroplasts caused by an in vivo excess of nickel. *Photosynthetica* **34**, 513-522.
- Margaret EF (1994) *Plants and Chemical Element*, pp 1-292. John Wiley & Sons, New York.
- Miranda MG and Hangovan K (1978) Uptake of boron by *Lemna minor*. *Aquat Bot* **4**, 53-64.
- Fargasova A (1999) The green alga *Scenedesmus quadricauda* – a subject for the study of inhibitory effects of Cd, Cu, Zn, Pb and Fe. *Biologia* **54**, 393-8.
- Lasheen MR (1990) Effect of cadmium, copper and chromium (VI) on the growth of Nile algae. *Water Air Soil Pollut* **50**, 19-30.
- Leborans GF, Novillo A (1996) Toxicity and bioaccumulation of cadmium in *Olishodiscus luteus* (Raphidophyceae). *Water*

- Res **30**, 57-62.
34. Pahlsson AMB (1989) Toxicity of heavy metals (Zn, Cd, Cu, Pb) to vascular plants. *Water Air Soil Pollut* **47**, 287-319.
 35. Sen AK and Mondal NG (1987) *Salvinia natan*-as the scavenger of Hg (II). *Water Air Soil Pollut* **34**, 439-46.
 36. Costa ACA and Leite SGF (1990) Cadmium and zinc biosorption by *Chlorella homosphaera*. *Biotechnol Lett* **12**, 941-44.
 37. Pawlik B (2000) Relationships between acid-soluble thiol peptides and accumulated Pb in the green alga *Stichococcus bacillaris*. *Aquat Toxicol* **50**, 221-30.
 38. Maine AM, Duarte MV and Sune NL (2001) Cadmium uptake by floating macrophytes. *Water Res* **35**, 2629-34.
 39. Wang TC, Weissman JC, Ramesh G, Varadarajan R and Benemann JR (1996) Parameters for removal of toxic heavy metals by water milfoil (*Myriophyllum spicatum*). *Bull Environ Contam Toxicol* **57**, 786-9.
 40. Matsunaga T, Takeyama H, Nakao T and Yamazawa A (1999) Screening of marine microalgae for bioremediation of cadmium-polluted seawater. *J Biotechnol* **70**, 33-8.
 41. Tokunaka T, Furuta N and Morimoto M (1976) Accumulation of cadmium in *Eichhornia crassipes*. *J Hyg Chem* **22**, 234-9.
 42. Zayed A, Gowthman S and Terry N (1998) Phytoaccumulation of trace elements by wetland plants : I. Duckweed. *J Environ Qual* **27**, 715-21.
 43. Garg P and Chandra P (1990) Toxicity and accumulation of chromium in *Ceratophyllum demersum* L. *Bull Environ Contam Toxicol* **44**, 473-8.
 44. Cain JR, Paschal DC and Hayden CM (1980) Toxicity and bioaccumulation of cadmium in colonial green alga *Scenedesmus obliquus*. *Arch Environ Contam Toxicol* **9**, 9-16.
 45. Rai UN and Chandra P (1992) Accumulation of copper, lead, manganese and iron by field population of *Hydrodictyon reticulatum* (Linn) Lagerheim. *Sci Total Environ* **116**, 203-11.
 46. Zhu YL, Zayed AM, Qian JH, Souza M and Terry N (1999) Phytoaccumulation of trace elements by wetland plants : II. Water hyacinth. *J Environ Qual* **28**, 339-44.
 47. Devi M, Thomas DA, Barber JT and Fingerham M (1996) Accumulation and physiological and biochemical effects of cadmium in a simple aquatic food chain. *Ecotoxicol Environ Saf* **33**, 38-43.
 48. Huebert DB and Shay M (1993) The response of *Lemna trisulca* L. to cadmium. *Environ Pollut* **80**, 247-53.
 50. Rai UN, Sinha S, Tripathi RD and Chandra P (1995) Wastewater treatability potential of some aquatic macrophytes : Removal of heavy metals. *Ecol Eng* **5**, 5-12.