

Andrographolide Ameliorates Beta-Naphthoflavone-Induced CYP1A Enzyme Activity and Lipid Peroxidation in Hamsters with Acute Opisthorchiasis

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Background: *Opisthorchis viverrini* (OV) infection generates oxidative stress/free radicals and is considered as a primary cause of cholangiocarcinoma since it primarily triggers sclerosing cholangitis.

Objective: In this study, the impacts of andrographolide on acute opisthorchiasis in β -naphthoflavone (BNF)-exposed hamsters were investigated.

Material and Method: Ethoxyresorufin O-deethylase (EROD) and methoxyresorufin O-demethylase (MROD) activities and Thiobarbituric acid reaction substances (TBARS) assay of andrographolide in acute opisthorchiasis in the BNF-exposed hamsters were assessed.

Results: The results showed that andrographolide ameliorated the hepatic CYP1A1 and CYP1A2 activities by decreases of the specific enzymatic reactions of EROD and MROD, respectively, in the BNF-exposed hamsters. Moreover, andrographolide lowered the formation of malondialdehyde in the livers and brains of the hamsters.

Conclusion: These observations revealed the promising chemo-protective and antioxidant activities of andrographolide via suppression of the specific EROD and MROD reactions and lipid peroxidation against acute opisthorchiasis in the BNF-exposed hamsters.

Keywords: Opisthorchiasis, Andrographolide, β -naphthoflavone, CYP1A, Lipid peroxidation, Hamster

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Infection of liver fluke or *Opisthorchis viverrini* (OV) is a major health problem in tropical countries of Southeast Asia⁽¹⁾. The excysted metacercariae then develop into juvenile flukes within the biliary tree of the liver, leading to acute cholangitis and develop into cholangiocarcinoma⁽²⁾. Besides the OV infection, cytochrome P450 1A (CYP1A) inducers such as the synthetic flavone, β -naphthoflavone (BNF), are known to induce oxidative stress and have a liver tumor-promoting effect in rats⁽³⁾. The oxidative stress is normally generated by microsomal electron systems including CYPs and nicotinamide adenine dinucleotide phosphate

(NADPH)-CYP reductase, leading to the production of reactive oxygen species (ROS) through metabolism processes, followed by a subsequent formation of oxygenated substrates and water⁽⁴⁾. CYP1A is the major enzyme involved in metabolic activation of a large number of procarcinogens to form reactive intermediates that can interact with cellular nucleophiles and ultimately trigger carcinogenesis⁽⁵⁾. The activities of CYP1A1 and CYP1A2 are associated with progression of various cancers⁽⁶⁾.

Andrographolide is the major diterpenoid constituent of *Andrographis paniculata*, a traditional medicine claimed for various pharmacological benefits⁽⁷⁾, i.e., hepatoprotective, anti-oxidative, anti-inflammatory, and anti-cancer activities, etc^(8,9). The diverse effects of *A. paniculata* extract or andrographolide on the expression of a limited number of CYPs in vivo in rat livers and in vitro in human and rat hepatocyte cultures have been noted⁽¹⁰⁾. Therefore,

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it is worth determining how andrographolide exhibits pharmacological actions against acute opisthorchiasis in the BNF-exposed hamsters. The present study aims to investigate chemo-protective and antioxidative effects of andrographolide in acute opisthorchiasis in the BNF-exposed hamsters via the assessment of EROD and MROD activities and thiobarbituric acid reactive substance (TBARS) assay. The results assured andrographolide as a promising chemo-protective candidate based on the significant suppression of the specific CYP1A1 and CYP1A2 enzymatic reactions and lipid peroxidation in the BNF-exposed hamsters with acute opisthorchiasis.

Material and Method

Materials

Andrographolide was a product of Wako Pure Chemical Industries (Osaka, Japan). BNF, ethoxyresorufin (ER), methoxyresorufin (MR), were products of Sigma Aldrich (St. Louis, MO, USA). All other laboratory chemicals were of the highest purity available from commercial suppliers.

Animals

Female Syrian Golden hamsters at 3 weeks of age were supplied by the Animal Unit of Faculty of Medicines (Khon Kaen University, Khon Kaen, Thailand). Animal handling and the treatment protocols were approved by the Animal Ethic Committee for use and care of Khon Kaen University, Thailand (Approval No. AEKKU01/2555). The hamsters were infected by a single intragastric gavage given 50 metacercaria of OV. Three weeks after the infection to OV, the hamsters were daily intra-peritoneally administered with BNF (30 mg/kg/day) for 3 days, and/or andrographolide (5 mg/kg/day) for 5 days. The livers and brains were collected by liver perfusion 24 h after the last treatment and immediately kept at -80°C for further analysis.

Preparation of hepatic microsome

The liver was weighed, minced, and homogenized on an ice-bath in three volumes of cold 1.15% (w/v) potassium chloride. The microsomal fraction was prepared by centrifugation as described⁽¹¹⁾.

Assessment of alkoxyresorufin O-dealkylase activity

Hepatic AROD activities were determined by the method of Jarukamjorn⁽¹¹⁾, with modifications.

Determination of lipid peroxidation

Lipid peroxidation was determined by

thiobarbituric acid fluorometric method⁽¹²⁾.

Statistical analysis

The results were expressed as means of triplicate independent duplicate experiments \pm SD and analyzed by using one-way analysis of variance (ANOVA) followed by Tukey post-hoc test (version 19; SPSS Inc., Chicago, IL, USA).

Results

Effect of andrographolide on hepatic EROD and MROD activities

The induction of EROD activity was noted in the acute opisthorchiatic hamsters (Fig. 1A). BNF1 significantly elevated the level of hepatic EROD activity in the acute OV-infected hamsters while andrographolide itself did not affect. Interestingly, andrographolide significantly restored the BNF-induced EROD activity to a similar level as that of the OV-infected hamsters, though it could not reach to the

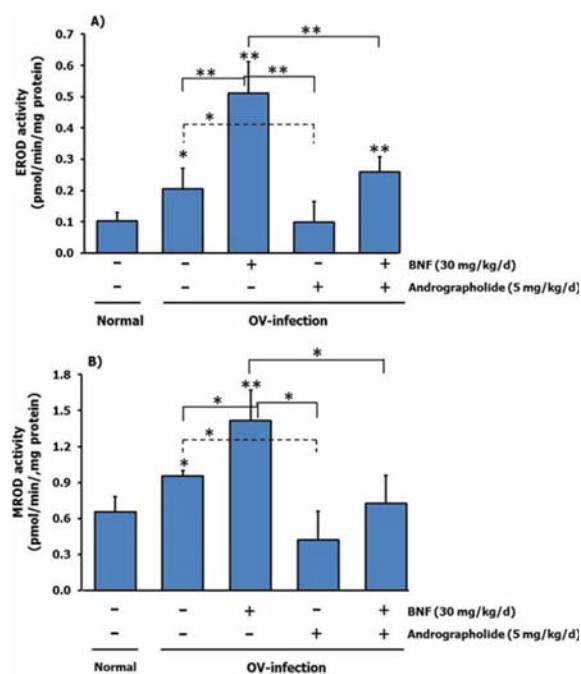


Fig. 1 EROD (A) and MROD (B) activities in the livers of female Syrian Golden hamsters. The hamsters were infected by a single intragastric administration of 50 OV metacercaria. Three weeks after the infection, the hamsters were daily administered with BNF (30 mg/kg/day) for 3 days, and/or andrographolide (5 mg/kg/day) for 5 days. The data is presented as the mean \pm SD (n = 3) from two independent experiments. * $p < 0.05$, ** $p < 0.01$.

baseline level of the non-treated hamsters. Besides the benefit of andrographolide on suppression of CYP1A1 in the BNF-exposed hamsters with acute opisthorchiasis, the BNF-induced MROD activity was returned to the baseline by andrographolide (Fig. 1B). Consistent with the EROD activity, BNF excessively increased the level of hepatic MROD activity in the acute OV-infected hamsters while andrographolide itself did not. These observations implied that andrographolide assures the chemo-protective effect by restoring the BNF-induction of CYP1A1 and CYP1A2 activity in the acute opisthorchiatic hamsters.

Effect of andrographolide on lipid peroxidation

The results from both liver and brain showed that the formation of MDA was significantly higher in either the OV-infected alone or the OV-infected with BNF-exposed hamsters (Fig. 2A&B). Likewise, andrographolide did not change the level of lipid peroxidation in the livers and brains of the OV-infected

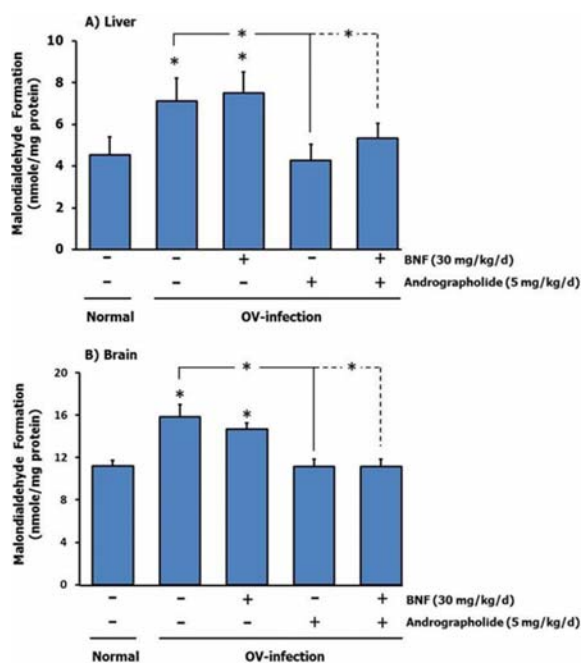


Fig. 2 Lipid peroxidation in the livers (A) and the brains (B) of female Syrian Golden hamsters. The hamsters were infected by a single intragastric administration of 50 OV metacercaria. Three weeks after the infection, the hamsters were daily administered with BNF (30 mg/kg/day) for 3 days, and/or andrographolide (5 mg/kg/day) for 5 days. The data is presented as the mean \pm SD (n = 3) from two independent experiments. * $p < 0.05$.

hamsters. Expressively, andrographolide significantly restored the level of lipid peroxidation in the BNF-exposed hamsters with acute opisthorchiasis to the same level as those of the controls.

Discussion

Infection with OV due to eating certain traditional freshwater-fish dishes is a principal risk factor for cholangiocarcinoma in the Northeast of Thailand where the infection is endemic and the incidence of this form of primary liver cancer has been the highest in the world⁽¹³⁾. During chronic inflammation, ROS and reactive nitrogen species are generated from inflammatory and epithelial cell and play a key role in pathologic conditions and carcinogenesis⁽¹⁴⁾. OV infection triggers inflammation surrounding the bile duct lumen in hamsters, and during acute inflammation, the oxidative/nitrative stress also induces inflammation liver injury⁽¹⁵⁾. The study of Pinlaor et al, 2004⁽¹⁵⁾ revealed that MDA level in the blood of hamsters fed with 50 metacercariae of OV by intragastric intubation for single infections was significantly increased from 3 to 90 days, agreed with our result. The MDA level was increased in the liver and brain of hamsters after 21 days of the OV infection. Andrographolide potentially returned the lipid peroxidation in both of the liver and brain of the acute opisthorchiasis hamsters. These observations corresponded to the previous study proposed that the antioxidant property of andrographolide could be due to activation of antioxidant enzymes⁽¹⁶⁾. Furthermore, the present findings supported the hepatoprotective property of andrographolide from previous studies in mice, rats, and guinea pigs⁽¹⁶⁻¹⁸⁾. The nuclear factor erythroid 2-related factor (Nrf2) is a transcriptional factor that functions as a key controller of the redox homeostatic gene-regulatory network⁽¹⁹⁾. Enhancement of oxidative stress and nitrative stress in the OV-infected hamsters may be mediated through the Nrf2 pathway by increasing expression of superoxide dismutase and Glutathione peroxidase enzymes (GPx), which suggested host defensive mechanisms against oxidative stress⁽²⁰⁾. Andrographolide was also found to be the most potent Nrf2 activator among 200 biologically active compounds⁽²¹⁾.

Until now, praziquantel has been widely used for the treatment of OV infection because of its apparent effectiveness⁽²²⁾. Praziquantel might produce general side effects, i.e., nausea, vomiting, headache, and abdominal discomfort⁽²³⁾. In addition, praziquantel treatment for short term induced inflammation, and

resulted in oxidative and nitrate stress through OV antigen release⁽²⁰⁾. Therefore, andrographolide might convey a benefit as an adjuvant of an anthelmintic agent because of its antioxidative property to reduce lipid peroxidation.

Besides the effect of ROS on cholangitis or cholangiocarcinoma, some hepatic metabolizing enzymes related cancer should be concerned. CYP1A1 and CYP1A2 play the role in metabolic activation of carcinogenic polycyclic aromatic hydrocarbons (PAHs) through aryl hydrocarbon receptor (AhR)⁽²⁴⁾. The implication of CYP1A induction in human risk evaluation has been concerned and remained a central focus of interest in cancer research, toxicology, and food safety⁽²⁴⁾. EROD and MROD activities were reduced in *Schistosomamansoni* infected DBA/2 mice⁽²⁵⁾. Furthermore, the activity of CYP1A1 and expression of CYP1A1/2 protein were significantly increased in the liver of *Taeniataeniformis* metacestodes infected rats⁽²⁶⁾. However, the effect of OV infection on expression of CYP1A was still unknown. This is the first time to report the induction of EROD and MROD in the OV-infected hamsters. BNF also additional increased the EROD and MROD activities. When people were infected with OV, they possibly exposed to procarcinogens in daily life via char-food and/or environmental pollutants, resulted in increasing cancer risk. Hence, it is of interest to have a dietary compound or natural produce to delay or lower the risk.

The very recent publication presented the anticancer potential of andrographolide in cholangiocarcinoma cells⁽²⁷⁾. These findings support that andrographolide possesses chemo-protective property with the ability to down-regulate CYP1A in the OV-infected hamsters exposed to a CYP1A inducer. Taken data together, andrographolide had chemo-protective property and anti-lipid peroxidation due to its ability to down-regulate both EROD and MROD activities, and MDA formation in the BNF exposed hamsters to acute opisthorchiasis.

This is the first time to report a powerful chemo-protective activity and antioxidant property of andrographolide via the specific responsible CYP1A1 and CYP1A2 enzymatic reactions, EROD and MROD, and formation of MDA, against the acute opisthorchiasis in BNF-exposed hamsters. Therefore, andrographolide might be a promising adjuvant against cholangiocarcinoma development in an OV-infected patient, at least via amelioration of CYP1A1/2-related carcinogenesis and oxidant-antioxidant balance

pathways.

What is already known on this topic?

Opisthorchis viverrini (OV) infection is considered as a primary cause of cholangiocarcinoma since it primarily triggers sclerosing cholangitis. A chronic infection of OV generates oxidative stress/free radicals and inflammation. Andrographolide is a Thai herbal medicine and has antioxidant property via activation of antioxidant enzymes. Taken data together, andrographolide might be a benefit to those who have OV infection.

What is this study adds?

The study implied that andrographolide has a chemo-protective effect by restoring the BNF-induction of CYP1A1 and CYP1A2 activity in the acute opisthorchiatic hamsters. Moreover, formation of MDA shows that andrographolide significantly restored the level of lipid peroxidation in the BNF-exposed hamsters with acute opisthorchiasis.

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Potential conflicts of interest

None.

References

1. Jattujan P, Pinlaor S, Charoensuk L, Arunyanart C, Welbat JU, Chaijaroonkhanarak W. Curcumin prevent bile canicular alterations in the liver of hamsters infected with *Opisthorchis viverrini*. Korean J Parasitol 2013; 51: 695-701.
2. Prasongwatana J, Laummaunwai P, Boonmars T, Pinlaor S. Viable metacercariae of *Opisthorchis viverrini* in northeastern Thai cyprinid fish dishes—as part of a rational program for control of *O. viverrini*-associated cholangiocarcinoma. Parasitol Res 2013; 112: 1323-7.
3. Dewa Y, Nishimura J, Muguruma M, Jin M, Saegusa Y, Okamura T, et al. beta-Naphthoflavone enhances oxidative stress responses and the induction of preneoplastic lesions in a diethylnitrosamine-

- initiated hepatocarcinogenesis model in partially hepatectomized rats. *Toxicology* 2008; 244: 179-89.
4. Poulos TL, Raag R. Cytochrome P450cam: crystallography, oxygen activation, and electron transfer. *FASEB J* 1992; 6: 674-9.
 5. Guengerich FP. Metabolism of chemical carcinogens. *Carcinogenesis* 2000; 21: 345-51.
 6. Kawajiri K, Nakachi K, Imai K, Watanabe J, Hayashi S. The CYP1A1 gene and cancer susceptibility. *Crit Rev Oncol Hematol* 1993; 14: 77-87.
 7. Jarukamjorn K, Nemoto N. Pharmacological aspects of *Andrographis paniculata* on health and its major diterpenoid constituent andrographolide. *J Health Sci* 2008; 54: 370-81.
 8. Jain PK, Khurana N, Pounikar Y, Gajbhiye A, Kharya MD. Enhancement of absorption and hepatoprotective potential through soya-phosphatidylcholine-andrographolide vesicular system. *J Liposome Res* 2013; 23: 110-8.
 9. Lim JC, Chan TK, Ng DS, Sagineedu SR, Stanslas J, Wong WS. Andrographolide and its analogues: versatile bioactive molecules for combating inflammation and cancer. *Clin Exp Pharmacol Physiol* 2012; 39: 300-10.
 10. Pekthong D, Martin H, Abadie C, Bonet A, Heyd B, Manton G, et al. Differential inhibition of rat and human hepatic cytochrome P450 by *Andrographis paniculata* extract and andrographolide. *J Ethnopharmacol* 2008; 115: 432-40.
 11. Jarukamjorn K, Sakuma T, Miyaura J, Nemoto N. Different regulation of the expression of mouse hepatic cytochrome P450 2B enzymes by glucocorticoid and phenobarbital. *Arch Biochem Biophys* 1999; 369: 89-99.
 12. Udomsuk L, Juengwatanatrakul T, Putalun W, Jarukamjorn K. Bimodal action of miroestrol and deoxymiroestrol, phytoestrogens from *Pueraria candollei* var. *mirifica*, on hepatic CYP2B9 and CYP1A2 expressions and antilipid peroxidation in mice. *Nutr Res* 2012; 32: 45-51.
 13. Saengsawang P, Promthet S, Bradshaw P. Infection with *Opisthorchis viverrini* and use of praziquantel among a working-age population in northeast Thailand. *Asian Pac J Cancer Prev* 2013; 14: 2963-6.
 14. Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002; 420: 860-7.
 15. Pinlaor S, Hiraku Y, Ma N, Yongvanit P, Semba R, Oikawa S, et al. Mechanism of NO-mediated oxidative and nitrative DNA damage in hamsters infected with *Opisthorchis viverrini*: a model of inflammation-mediated carcinogenesis. *Nitric Oxide* 2004; 11: 175-83.
 16. Trivedi NP, Rawal UM, Patel BP. Hepatoprotective effect of andrographolide against hexachloro-cyclohexane-induced oxidative injury. *Integr Cancer Ther* 2007; 6: 271-80.
 17. Roy DN, Sen G, Chowdhury KD, Biswas T. Combination therapy with andrographolide and d-penicillamine enhanced therapeutic advantage over monotherapy with d-penicillamine in attenuating fibrogenic response and cell death in the periportal zone of liver in rats during copper toxicosis. *Toxicol Appl Pharmacol* 2011; 250: 54-68.
 18. Shukla B, Visen PK, Patnaik GK, Dhawan BN. Choleretic effect of andrographolide in rats and guinea pigs. *Planta Med* 1992; 58: 146-9.
 19. Yu AL, Lu CY, Wang TS, Tsai CW, Liu KL, Cheng YP, et al. Induction of heme oxygenase 1 and inhibition of tumor necrosis factor alpha-induced intercellular adhesion molecule expression by andrographolide in EA.hy926 cells. *J Agric Food Chem* 2010; 58: 7641-8.
 20. Pinlaor S, Prakobwong S, Hiraku Y, Kaewsamut B, Dechakhamphu S, Boonmars T, et al. Oxidative and nitrative stress in *Opisthorchis viverrini*-infected hamsters: an indirect effect after praziquantel treatment. *Am J Trop Med Hyg* 2008; 78: 564-73.
 21. Smirnova NA, Haskew-Layton RE, Basso M, Hushpilian DM, Payappilly JB, Speer RE, et al. Development of Nrf2-luciferase reporter and its application for high throughput screening and real-time monitoring of Nrf2 activators. *Chem Biol* 2011; 18: 752-65.
 22. Boonmars T, Srisawangwong T, Srirach P, Kaewsamut B, Pinlaor S, Sithithaworn P. Apoptosis-related gene expressions in hamsters re-infected with *Opisthorchis viverrini* and re-treated with praziquantel. *Parasitol Res* 2007; 102: 57-62.
 23. Mairiang E, Mairiang P. Clinical manifestation of opisthorchiasis and treatment. *Acta Trop* 2003; 88: 221-7.
 24. Ma Q, Lu AY. CYP1A induction and human risk assessment: an evolving tale of in vitro and in vivo studies. *Drug Metab Dispos* 2007; 35: 1009-16.
 25. Conte FP, Fidalgo-Neto AA, Manhaes-Rocha DA, Paumgartten FJ, De Oliveira AC. Activity of liver

- microsomal enzymes during the chronic phase of murine schistosomiasis. *Braz J Med Biol Res* 2007; 40: 657-62.
26. Montero R, Serrano L, Davila VM, Ito A, Plancarte A. Infection of rats with *Taenia taeniformis* metacestodes increases hepatic CYP450, induces the activity of CYP1A1, CYP2B1 and COH isoforms and increases the genotoxicity of the procarcinogens benzo[a]pyrene, cyclophosphamide and aflatoxin B(1). *Mutagenesis* 2003; 18: 211-6.
27. Suriyo T, Pholphana N, Rangkadilok N, Thiantanawat A, Watcharasit P, Satayavivad J. *Andrographis paniculata* extracts and major constituent diterpenoids inhibit growth of intrahepatic cholangiocarcinoma cells by inducing cell cycle arrest and apoptosis. *Planta Med* 2014; 80: 533-43.

ผลของสารแอนโดรกราโฟไลด์ต่อสมรรถนะของเอนไซม์ CYP1A ในภาวะที่ถูกกระตุ้นด้วยสารบีตาแนพโทฟลาโวน และภาวะ
 ลิปิดเปอร์ออกซิเดชันในหนูแฮมสเตอร์ที่เกิดภาวะ opisthorchiasis แบบเฉียบพลัน

ลติพร อุดมสุข, วรรษญา จตุพรประเสริฐ, กนกวรรณ จารุกำจร, ไพบุลย์ สิทธิถาวร

ภูมิหลัง: การติดเชื้อพยาธิใบไม้ตับ (*Opisthorchis viverrini*) ทำให้เกิดภาวะเครียดออกซิเดชันและอนุมูลอิสระ และถือเป็นสาเหตุสำคัญที่ทำให้เกิด
 ภาวะตับอ่อนอักเสบและนำไปสู่โรคมะเร็งถุงน้ำดี

วัตถุประสงค์: การศึกษาในครั้งนี้จึงมุ่งเน้นศึกษาผลของสารแอนโดรกราโฟไลด์ต่อการเกิดภาวะ opisthorchiasis แบบเฉียบพลันในหนูแฮมสเตอร์
 ที่ได้รับสารบีตาแนพโทฟลาโวน

วัสดุและวิธีการ: ทดสมรรถนะของเอนไซม์ ethoxyresorufin O-deethylase (EROD) และ methoxyresorufin O-demethylase (MROD)
 และใช้วิธี TBARS ของสารแอนโดรกราโฟไลด์ในแฮมสเตอร์ที่ได้รับสารบีตาแนพโทฟลาโวนและเกิดภาวะ opisthorchiasis แบบเฉียบพลัน

ผลการศึกษา: ผลการทดลองพบว่าสารแอนโดรกราโฟไลด์สามารถลดสมรรถนะของเอนไซม์ CYP1A1 และ CYP1A2 ผ่านปฏิกิริยา EROD และ
 MROD ในแฮมสเตอร์ที่ได้รับสารบีตาแนพโทฟลาโวน นอกจากนี้สารแอนโดรกราโฟไลด์ยังสามารถลดการสร้างสารมาลอนไดอัลดีไฮด์ในตับและสมอง
 ของแฮมสเตอร์อีกด้วย

สรุป: จากผลการทดลองดังกล่าวแสดงให้เห็นถึงศักยภาพของสารแอนโดรกราโฟไลด์ในการเป็นสารต้านมะเร็ง และต้านอนุมูลอิสระผ่านการลด
 การทำงานของปฏิกิริยา EROD และ MROD และลดการเกิดลิปิดเปอร์ออกซิเดชัน ในหนูแฮมสเตอร์ที่ได้รับสารบีตาแนพโทฟลาโวนที่มีภาวะ opisthor-
 chiasis แบบเฉียบพลัน
