The Roles of Calcium Lactate on Bruised Damage in Papaya Fruit

Khurnpoon, L. 1* and Siriphanich, J. 2

¹Department of Plant Production Technology, Faculty of Agricultural Technology, King Mongkut's Institute of Technology Ladkrabang, Bangkok 10520, Thailand; ²Department of Horticulture, Faculty of Agriculture, Kasetsart University, Kamphaeng Saen Campus, Nakon Pathom 73140, Thailand.

Khurnpoon, L. and Siriphanich, J. (2018). The roles of calcium lactate on bruised damage in papaya fruit. International Journal of Agricultural Technology 14(6):871-880.

Abstract Bruise damage reduces postharvest quality and it was rejected by consumer. Lack detail about bruise mechanism reported in papaya fruit, the prevention and reduction of postharvest loss were limited. The tissue softening as a result in the change of cell wall solubility and depolymerization including changes in physical properties were following bruise damaged. The maintaining cell wall integrity by calcium lactate application before bruising was tested. Papaya fruits were non-dipped or dipped in 2% calcium lactate solution at 55°C for 5 minutes, then air dried at room temperature for 1 hour. Two groups of fruit sample were separated; one group was impacted from height level at 75 cm while the other was nonimpacted. The result showed that bruise treatment reduced skin lightness, chroma and hue angle especially in 'Khak dam' papaya, but could maintain by calcium lactate application. This treatment also preserved the reduction of fruit firmness by 4 days concomitant with the solubilization of pectin during bruising was reduced by calcium lactate treatment. Water-, Na₂CO₃ - and potassium hydroxide-soluble fraction were low in calcium lactate treatment both with and without bruise damage. Non-significant different between non-dipped and dipped in calcium lactate in TSS, skin color development and fresh weight loss in bruise treatment. However, calcium lactate treatment reduced the increase in bruise area of 20 and 10% in 'Khak dam' and 'Holland' papaya, respectively. Fruit decay incidence was 5% low in fruit treated with calcium lactate following bruise damage than non-dipped.

Keywords: calcium lactate, bruise, pectin, cell wall, softening

Introduction

Bruise damage had been report to reduce fruit quality in many fruits (Ahmadi, 2010). The symptom mainly skin browning and pulp softening increase within 2 hours after impact in tomato, apple and pear (Van linden *et al.*, 2008; Garcia *et al.*, 1995; Blahovee and Paprstein, 2005). In papaya, bruise severity differ among cultivars generally more severe in 'Khak dum' than

^{*}Corresponding Author: Khurnpoon, L.; Email: kplampan@gmail.com

'Holland' papaya (Khurnpoon and Siriphanich, 2011) as a results of different in cell wall pectin component and molecular size distribution of pectin where changed to smaller size during storage of bruised fruit (Goulao et al., 2008). Calcium solution mostly as calcium chloride and calcium lactate play a role to preserve the integrity of cell wall resulted in delay fruit softening. The firming preservation by calcium solution may build a complexion of calcium ions with cell wall and middle lamella pectin (Morris, 1980) that helps to stabilize of cell wall and effect of cell turgor pressure (Mignani, 1995). However, the unfavorable was affect by using calcium chloride for texture preservation (Monsalve-Gonzalez et al., 1993) and appeared the residual amount of calcium choloride remained on the surface of the products that was not accepted by the consumer. Calcium lactate was represents an alternative calcium source without reported flavor difference. Calcium lactate at concentration between 0.5-2.0% has been used as a firming agent for processed strawberry (Morris et al., 1985). and grapefruit (Baker, 1993). In addition, the application of calcium lactate has been reported in some fresh cut produces. Application of 2.0% of calcium lactate helps to preserve pulp firmness and delayed the changes in fruit quality during storage at ambient temperature for 70 days (Rehman et al., 2017). Many reports showed that the combined use of calcium lactate with heat treatment gave the better results of firming preservation in fresh cut apple (Alandes et al., 2006), carrot (Rico, 2007) and lettuce (Martin-Diana, 2006). Limited information exists on the effect of bruise damage on cell wall metabolism particularly pectic component of the primary cell wall. Increased pectin degradation was found in wounded tissue of fresh cut tomato which causes by pectin hydrolysis as a resulted in wound-induced enzyme synthesis (Huber et al., 2001). In contrast with the compared study between intact and fresh cut tomato by Chung et al. (2006) found limited activities in the cut fruit. However, the associated change in the cell wall pectin was not examined. Beside the change in cell wall pectin, the change in xyloglucan network was likely to be associated with bruising (Hadfield and Bennett, 1998). The results in tomato showed no substantial change in PME and PG activity with bruising, although PG activity increased significantly with ripening. The degree of demethoxylation was slightly reduced in wounded tissue 3 h after impactbruising. Bruising did not lead to significant changes in pectin solubility or degree of polymerization within 3 h of impact (Van linden et al., 2008). Although calcium solution acts as firming agent in many kinds of fruits, the effect of calcium lactate application before mechanical impact bruising in papaya has not been studied yet. Our study aims to understand the effect of calcium lactate on bruise damage of papaya fruit. The work focused on the change in physiological and biological of cell wall pectin.

Materials and methods

'Khak dum' and 'Holland' papaya were harvested at maturity stage (25 and 50% of yellow skin color developed) with uniform shape and size. Fruit samples were washed with chlorinated water then air dried at room temperature. A total of 72 fruits for each cultivar were obtained for 6 treatments. Fruit samples were dipped in water (control treatment) or 2% (w/w) calcium lactate solution for 5 min in 55°C then placed on plastic tray until dried and stored at room temperature for 24 h before bruising. Mechanical impact bruising for bruise treatments were obtained by dropped from height level at 75 cm. Impact site was marked with permanent-ink and started to evaluated physiological changes after 2 h of impact that was represented for day 0. The experiment was conducted using a completely randomized design (CRD) and comparison of treatment means was analyzed using Duncan's multiple range test (DMRT) at 5% level. Fruit physiological parameters and Cell wall extraction and analysis were recorded as followed.

Change in fruit skin color

Skin color development was determined by using the Color flex spectrophotometer. Data were reported as L value (lightness) ranging from 0 (brightness) -100 (darkness), chroma (C) and hue angle (H) values were represented to the saturated and the purity of color, respectively.

Change in fruit quality

The position of damage areas for each fruit sample after dropped from height level at 75 cm was labelled with permanent ink. The area of damage was evaluated by drawing an area on tracing paper then transferred to a graph paper in the area size of about 1 square centimeter. The damage area was reported as square centimeter (cm²). Change in fruit firmness was measured by fruit firmness tester with a 5 mm diameter of plunger head. Fruit samples were cut in cross section for 2 pieces each then press the plunger head depth approximately 0.5 cm. The data was reported in Newton (N). The total soluble solids (TSS) content was determined from blended 10g of pulp tissue to get about 5 ml of same juice. Two drop of fruit juice was dropped on prism of hand refractometer (Effigi®) then read through an eye piece and reportrf in the unit of %brix. After determined all quality parameters, the tissue samples were collected from the damage area. The sample was cut into small pieces then kept in freezers at -20°C for cell wall extraction and analysis.

Cell wall extraction and analysis

An alcohol insoluble residue (AIR) was prepared from frozen tissue by thawing and left 1 h prior to extraction. Approximately 20 g of tissue was boiled in 100 ml ethanol at 70°C for 5 minutes then blended and filtered through whatman#1 filter paper. The residues were washed with excess alcohol and acetone, respectively. The residues were then air dried with remained constant in weight. Dry samples as represented AIR were ground in mortar and stored in desiccators until analysis. AIR of 50 mg was weight and extracted with 20 ml distilled water in 50 ml Erlenmeyer flash at room temperature for 2 hours with shaking. The suspension was then centrifuged at 6,000g for 15 minutes and the supernatant was collected and kept in plastic conical tube at 15°C for analyze. The remained residues were re-extracted with the same amount of distilled water as described previous above. The supernatant were pooled with the earlier collected and represented as water soluble fraction (WSF). The residues from WSF was then sequentially extracted with 50 mM CDTA containing 100 mM potassium acetate pH 4.0, 50 mM Na₂CO₃ containing 20 mM NaBH4 and 4 N KOH, respectively. Each fraction was done twice in the same amount of extractant and same steps as descript for WSF. The supernatants were represented as CDTA-soluble fraction (CSF), Na₂CO₃soluble fraction (NSF) and KOH-soluble fraction (KSF), respectively. The uronic acid content (µg galacturonic acid/mg AIR) was analyzed in triplicate by following the method of Blumenkrantz and Asboe-Hansen (1973) with slightly modified.

Statistical analysis

This experiment was designed as completely randomized designed (CRD). All treatments were run with at six replicates. Data was comparisons by analysis of variance (ANOVA) followed by Duncan's multiple range test with significance level at p<0.05.

Results

Fruit skin lightness in 'Khak dam' and 'Holland' papaya increased slowly during the first 2 days. The reduction of skin lightness was found in the fruit impacted from height level at 75 cm whereas continue increased in control of 'Khak dam' papaya. In 'Holland' papaya, the skin lightness for control and bruised treatment slightly increased until days 4 then declined until days 6. Skin lightness and chroma value showed significantly different between bruised

treatment and other but only in 'Khak dam' papaya. No significantly different had found in all color indices; lightness, chroma and hue angle values after 6 days in storage for 'Holland' papaya (Table 1).

Percentage of bruised area incidence appeared in fruit skin of 'Khak dam' papaya 2 days earlier than 'Holland' papaya. Bruise severity as showed by bruise area on the last day in storage, approximately 25 and 12% in 'Khak dam' and 'Holland' papaya, respectively (Figure 1A and 1B).

Pulp firmness in 'Khak dam' papaya rapidly decreased in pulp firmness by 40-120 N from the beginning. Calcium lactate at 2% reduced the softening of fruit pulp by 25 and 15 N as compared to bruised and non-bruised treatment without calcium lactate application. However, the pulp firmness continuously decreased through the end of storage, ranged from 4.89-12.62 N. In 'Holland' papaya, fruit firmness decreased rapidly on days 2, about 20-120 N from the beginning. Bruise treatment enhanced the reduction of pulp firmness by 40 N as compared to non-bruised while treatment of 2% calcium lactate could preserved the pulp firmness by 43 when compared to calcium treated with and without bruised damage. On days 6, the highest in pulp firmness was found in control and 2% calcium lactate treatments, approximately 15 N higher than bruised treatment without calcium lactate application (Figure 2A and 2B).

Table 1. Skin lightness, chroma and hue angle values change in control (non-bruised) and bruised papaya with and without calcium lactate application after 6 days of impact.

| Cultivars | Treatments | Skin color parameters | | | | |
|-----------|------------------|-----------------------|-------------|---------------|--|--|
| | | Lightness | Chroma | Hue | | |
| Khak | Control | 44.77±3.12a | 32.27±3.12a | 1.49±0.26 | | |
| dam | Bruised | 37.89±3.39b | 18.70±1.77c | 1.27 ± 0.41 | | |
| | 2%Ca+Non-bruised | 46.24±2.31a | 32.96±2.39a | 1.40 ± 0.38 | | |
| | 2%Ca+Bruised | 45.49±2.54a | 29.20±2.55b | 1.39 ± 0.33 | | |
| | F-test | * | * | ns | | |
| Holland | Control | 58.72±3.91 | 52.31±3.22 | 1.10±0.43 | | |
| | Bruised | 57.50±2.77 | 55.24±2.95 | 1.11 ± 0.27 | | |
| | 2%Ca+Non-bruised | 56.12±2.85 | 50.40±3.07 | 1.07 ± 0.33 | | |
| | 2%Ca+Bruised | 56.61±3.04 | 48.66±4.65 | 1.08 ± 0.32 | | |
| | F-test | ns | ns | ns | | |

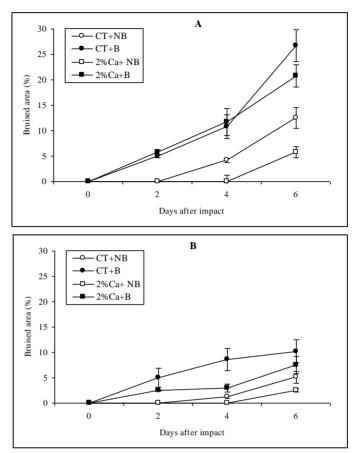


Figure 1. Percentage of bruised area in 'Khak dam' (A) and 'Holland' (B) papaya fruit after dipped and non-dipped in 2% calcium lactate solution then impacted from height level at 75 cm during 6 days in storage at room temperature.

No significantly different among treatment in total soluble solids content, slightly increased from 8 to 12 % brix on day 2 then declined to the same content of the beginning in 'Khak dam' papaya. Otherwise, TSS content remained constant at 12 % brix during 4 days after impacted with slightly increased in 75 cm of impacted height without calcium lactate application from 11.8 to 12.8 % brix then declined. Calcium application could reduce the increase in fresh weight loss during storage duration by 5 and 10% in 'Khak dam' and 'Holland' papaya, respectively (Table 2).

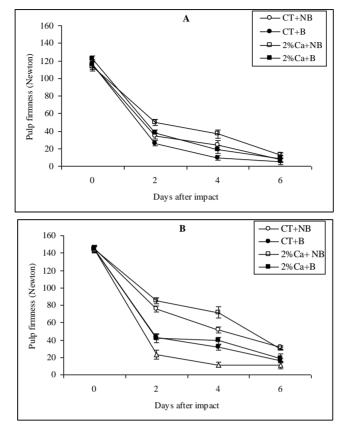


Figure 2. Pulp firmness in 'Khak dam' (A) and 'Holland' (B) papaya fruit after dipped and non-dipped in 2% calcium lactate solution then impacted from height level at 75 cm during 6 days in storage at room temperature.

Uronic acid content in 'Khak dam' and 'Holland' papaya had more solubilized in WSF than other fractions. On 6 days after impacted, bruise treatment showed high WSF content by 470.17 and 489.40 µg galacturonic acid/mg AIR in 'Khak dam' and 'Holland' papaya, respectively. The lowest WSF content was found in fruit treated with 2% calcium lactate without bruised impact by 281.83 and 371 µg galacturonic acid/mg AIR in 'Khak dam' and 'Holland' papaya, respectively. CSF content appeared to be high in bruised treatment in 'Khak dam' while it was lower in fruit treated with 2% calcium lactate without bruised damage treatment. Fruit treated with 2% calcium lactate then impacted from height level of 75 cm had higher in uronic acid content, at about 326.46 and 260.70 µg galacturonic acid/mg AIR in 'Khak dam' and 'Holland' papaya, respectively. NSF and KSF showed lowest content in fruit treated with 2% calcium lactate then impact from height level of 75 cm. 'Khak

dam' papaya had more solubilized of cell wall pectin in NSF and KSF approximately 4 and 1.5 times than 'Holland' papaya (Table 3).

Table 3. Uronic acid content (µg galacturonic acid/mg AIR) in the different pectin fractions of control (non-bruised) and bruised papaya tissue with and without calcium lactate application after 6 days of impact.

| | | 1 1 | <i>J</i> | 1 | | | |
|-----------|--------------|---|----------------|----------------|----------------|--|--|
| Cultivars | Treatments | Uronic acid content (µg galacturonic acid/mg AIR) | | | | | |
| | | WSF | CSF | NSF | KSF | | |
| Khak | Control | 409.94±14.21b | 160.09±15.06c | 144.72±12.05a | 160.09±15.62ab | | |
| dam | Bruised | 470.17±19.27a | 199.22±14.33b | 148.20±20.77a | 170.52±19.88a | | |
| | 2%Ca+Non- | 281.83±26.33d | 229.65±11.31a | 133.13±15.64b | 152.26±13.36b | | |
| | bruised | | | | | | |
| | 2%Ca+Bruised | 326.99±31.72c | 216.46±10.09a | 94.58±10.89c | 129.65±12.42c | | |
| | F-test | * | * | * | * | | |
| Holland | Control | 439.80±20.11b | 140.10±15.39c | 169.90±10.92a | 195.40±26.14a | | |
| | Bruised | 489.40±15.63a | 126.20±17.16c | 152.60±15.38 b | 153.40±18.33b | | |
| | 2%Ca+Non- | 371.10±17.58d | 386.40±16.03a | 32.80±7.81c | 53.40±16.59d | | |
| | bruised | | | | | | |
| | 2%Ca+Bruised | 385.90±20.52c | 260.70±20.48 b | 22.10±4.39c | 86.50±10.31c | | |
| | F-test | * | * | * | * | | |

Water soluble fraction (WSF), chelator soluble fraction (CSF), carbonate soluble fraction (NSF) and potassium hydroxide fraction (KSF)

Discussion

Fruit damage became soften with 2 days, about 5 times softer than the beginning. This change was parallel to the decrease in the skin lightness especially at the impact side. The bruise area appeared not much change to the lesser skin lightness as report in apple (Mitsuhashi-Gonzalez et al., 2010). It could explain by the different in major chemical composition between papaya and apple. High phenolic content in apple much appeared the skin or flesh browning after impact. The high calcium concentration resulted in decrease of fresh browning symptom. This symptoms have been directly associated with calcium content in many kinds of fresh. Therefore, calcium dips raise the possibility of producing fruits less susceptible to fresh browning symptoms. The same action of calcium salts has also been reported for fresh-cut fruit (Luna-Guzman and Barrett, 2000), where the enzymatic browning of fresh is result of different metabolic pathways. However, bruise damage affected the development of skin color (yellowness) by having non uniformity in 'Khak dam' papaya but not in 'Holland' papaya. This mechanical damage also caused an abnormal or uneven ripening in papaya. The mind effect of calcium lactate was found from this study by maintaining the reduction of skin lightness in 'Khak dam' papaya. The fruit quality as report by TSS content was not influenced by the postharvest calcium dips, and had no affect by bruise impacted. The effect of calcium lactate on fruit quality was found only on fruit softening. Calcium lactate application at 2% reduces the decrease in pulp firmness both in 'Khak dam' and 'Holland' papaya. The results of cell wall solubilization markedly in WSF showed the positive correlation with the reduction in pulp softening. This could explain by the change in pectin properties from protopectin to be a soluble form. Postharvest calcium application could maintain cell turgor, membrane integrity, tissue firmness and delays membrane lipid catabolism, extending storage life of fresh fruits (Garcia et al., 1996). Chardonnet et al. (2003) reported that 2% calcium chloride was enough to maximum calcium accumulation in the cell wall. Calcium links with pectin to form of 'egg box' model leading to cell wall network that increase wall strength (White and Broadley, 2003).

Acknowledgement

The author would like to thanks the Thailand Research Fund for financial support.

References

- Alandes, L., Hernado, I., Quiiles, A., Perez-Muauera, I. and Luach, M. A. (2006). Cell wall stability of fresh-cut Fuji apples treated with calcium lactate. Journal of Food Science. 71:615-620.
- Ahmadi, E., Ghassemzadeh, H.R., Sadeghi, M., Moghaddam, M. and Neshat, S. Z. (2010). The effect of impact and fruit properties on the bruising of peach. Journal of Food Engineering. 97:110-117.
- Alandes, L., Hernando, I., Quiles, A., Pérez-Munuera, I., Lluch, M. A. (2006). Cell wall stability of fresh-cut Fuji apples treated with calcium lactate. J. Food Sci. 71: 615-620.
- Blahovee, J. and Paprstein, F. (2005). Susceptibility of pear varieties to bruising. Postharvest Biology and Technology. 38:231-238.
- Chardonnet, C. O., Charron, C. S., Sams, C. E. and Conway, W. S. (2003). Chemical changes in the cortical tissue and cell walls of calcium infiltrated 'Golden Delicious' apples during storage. Postharvest Biology and Technology. 28:97-111.
- Chung, T.T., West, G. and Tucker, G. A. (2006). Effect of wounding on cell wall hydrolase activity in tomato fruit. Postharvest Biology and Technology. 40:250-255.
- Blumenkrantz, N. and Asboe-Hansen, G. (1973). New method for quantitative determination of uronic acids. Analytical Biochemistry. 54:484-489.
- Garcia, J.L., Ruiz-Altisen, M. and Barreiro, P. (1995). Factors influencing mechanical properties and bruise susceptibility of apples and pears. Journal of Agrictulture and Engineering Research. 61:11-18.
- Garcia, J. M., Gutierrez, F., Castellano, J. M., Perdiguero, S. and Albi, M. A. (1996). Influence of storage temperature on fruit ripening and olive oil quality. Journal of Agricultural and Food Chemistry. 44:264-267.
- Goulao, L. F. and Oliveira, C. M. (2008). Cell wall modifications during fruit ripening: when a fruit is not the fruit. Trends in Food Science and Technology. 19:4-25.

- Hadfield, K. A. and Bennett, A. B. (1998). Polygalacturonases: many genes in search of a function. Plant Physiology. 117:337-343
- Huber, D. J., Karakurt ,Y. and Jeong, J. (2001). Pectin degradation in ripening and wounded fruits. Brazilian Journal of Plant Physiology. 13:224-241.
- Khurnpoon, L. and Siriphanich, J. (2012). Change in fruit quality and cell wall polysaccharides in bruised papaya cultivar 'Khak Dum' and 'Holland'. Acta Horticulturae. 945:381-389.
- Luna-Guzman, I. and Barette, D. M. (2000). Comparison of calcium chloride and calcium lactate effectiveness in maintaining shelf stability and quality of fresh-cut cantaloupe. Postharvest Biology and Technology. 19:61-72.
- Martin-Diana, A. B., Rico, D., Firas, J. M., Henechan, G. T. M., Mulcahy, J., Barat, J. M. and Barry-Ryan, C. (2006). Effect of calcium lactate and heat-shock on texture in fresh-cut lettuce during storage. Postharvest Biology and Technology. 77:1069-1077.
- Mignani, I., Greve, L.C., Ben-Arie, R., Stotz, H.U., Li, C., Shakel, K. and Labavitch, J. (1995). The effect of GA, and divalent cations on aspects of pectin metabolism and tissue softening in ripening tomato pericarp. Physiology of Plant. 93:108-115
- Mitsuhashi-Gonzalez, K., Pitts, M. J., Fellman, J. K., Curry, E. A. and Clary, C. D. (2010). Bruising profile of fresh apples associated with tissue type and structure. Applied Engineering in Agriculture. 26:509-517.
- Monsalve-Gonzalez, A., Barbosa-Canovas, G. V. and Cavalieri, R. P. (1993). Mass transfer and textural changes during processing of apples by combined methods. Journal of Food Science. 58:1118-1124
- Morris, J. R., Sistrunk, W. A., Sims, C. A. and Main, G. L. (1985). Effect of cultivar, postharvest storage, preprocessing dip treatment and style of pack on the processing quality of strawberries. Journal of American Society for Horticultural Science. 110:172-177.
- Rehman, A. U., Ali1, A., Khan1, A., Qayum, A., Rahman, Z., Waqas, H., and Rehman, Z. U. (2017). Effect of calcium lactate concentrations on post-harvest storage life of apple fruit. Pure and Applied Biology. 6:1340-1344.
- Rico, D., Martin-Diana, A. B., Firas, J. M., Barat, J. M., Henechan, G. T. M. and Barry-Ryan, C. (2007). Improvement in texture using calcium lactate and heat-shock treatments for stored ready-to-eat carrots. Journal of Food Engineering, 79:1196-1206
- Van linden, V., Sila, D. N., Duvetter, T., De Baerdemaeker, J. and Hendrickx, M. (2008). Effect of mechanical impact-bruising on polygalacturonase and pectinmethylesterase activity and pectic cell wall components in tomato fruit. Postharvest Biology and Technology. 47:97-106.
- White, P. J. and Broadley, M. R. (2003). Calcium in plants. Annals of Botany. 92:487-511.

(Received: 23 July 2018, accepted: 30 October 2018)