Isolation and Pathogenicity of the Fungus, *Fusarium Solani* A Causal of Dry Root Rot on Sour Orange In Baghdad Province, Iraq

Mazin H. H. Al-Karboli 1 and Wadeja M. Kuthair 2

Department of Plant Protection, College of Agriculture, University of Baghdad, Iraq

Al-Karboli and Kuthair. (2016). Isolation and pathogenicity of the Fungus, *Fusarium solani* a causal of dry root rot on sour orange in Baghdad province. International Journal of Agricultural Technology 12(5):927-938.

Laboratory and field experiment were conducted at the college of Agriculture, University of Baghdad to investigate the variability in cultural, Morphological characteristic and pathogenicity of the casual of dry root rot, *Fusarium solani* on sour orange from different regions in Baghdad province. Results indicated the presence of 16 *F. solani* isolates and the appearance of the fungus at high rates ranged between (93-100) percent in all regions. It might be the dominant fungus in the roots of the sour orange .There were a significant differences between these isolate, Pathogenicity tests of these isolates on cabbage and sour orange seeds indicated that the percentage of germination between (3.3-26.3) % and (30-70) % for cabbage seed and sour orange respectively compared with 96.66 % for the control treatment. Al-Jadriyah isolates considered the most effective and the high pathogenicity where sour orange seed germination did not exceed 38.3%, while it ranged from (48.3-66.6) % for the rest of other isolates.

Keywords: - Fusarium solani isolates, Pathogenicity, dry root rot, sour orange, Iraq.

Introduction

Fusarium solani a ubiquitous fungus in the soil and affects a large number of crops, vegetables, trees, causing various symptoms which include: seed rotting, death and damping off seedlings, dry root rot (Kunta, et al.,2015; Domsch, et al.,1980; Farr, et al.,1989; Brasileiro et al., 2004). Fusarium solani mainly infects citrus trees especially grape fruit, citrus paradise, orange C. cinensis and to a lesser extent lemon, C. limon and trifoliate orange. Fusarium spp. Were isolated for the first time from citrus roots by Sherbakoff (1953). Studies on the roles of F. solani on citrus trees indicated a

^{*}Corresponding author: Mazin H. H. Al-Karboli E-mail: Mazinh91@gmail.com

significant reduction in seedlings growth and dry weight by 33%. Apparently, leaves of healthy citrus trees, suddenly wilt and turn yellow and die, roots are blackened and rotted with brown, vascular discoloration with the stem of the rootstock, fruits dry and stay on the tree (Olsen, *et al.*,2000).

Tatum and Baker (1983) explain the ability of *F. solani* to produce 11 form of toxins belong to naphthazarin group. And that some of these toxin can induce disease symptoms in citrus seedling represented by veins coloration, leaves wilting and clogged of transporting vessel. Also there is a relation between *Fusarium solani* races and it virulence and ability for toxin production and speed of disease symptoms appearance (Kumar, *et al.*, 1995; Baker, *et al.*, 1981). In nature plant pathogens exist as different strains that exhibit variation in their morphological and cultural characters, Pathogenicity and virulence (Ravi and Kumar, 2012). Morphological and Pathogenic variations are known in many fungal pathogens (Kumar, *et al.*, 1995; Khosrow and Zakaria, 2015; Kunta, *et al.*, 2015).

Dry root rot is considered is important disease on citrus in Iraq and there is no details studies about it, The present study was conducted to investigate the variability in cultural and morphological characteristic's and pathogenicity of *F. solani* isolates from different areas of sour orange orchards in Baghdad province

Material and Methods

Isolation and Identification of F. solani from sour orange roots

Isolates of F. solani were collected from 8 sour orange around Baghdad province Table (1) during the period of September – December 2015. 5 sample of roots were taken randomly from 5 trees showed symptom of common dry root rot in each orchard, then roots were transformed to lab and put in the refrigerator until isolation, and rinsed in tap water for 30-60 min, cut the roots into small pieces length of 0.5 - 1 cm showing vascular discoloration were surface sterilization for 3 minutes in sodium hiboklorate solution (1% free chlorine) and then washed with sterile water and dried with tissue paper. 4 sterile root pieces were cultured in petri dishes (9 cm diameter) contain 15-20 ml of potato dextrose agar (PDA). All culture plates were incubated in darkness at 25 ± 1 for 5 days. Fungal colonies resembling F. solani were identified depending on their cultural and characteristics using method described by

(Booth, 1977). F. solani were subcultured again and purified by using single spore technique (Salam, 2009) isolates were also grown on PDA (Nelson, et al., 1983) for the development of color.

The percentage of the appearance of *F. solani* in the root pieces were calculated from the equation of (Booth, 1977) below:

% of appearance
$$F$$
. $solani =$

Total numbers of pieces used for each sample

Pathogenicity of F. solani isolates on cabbage seeds

Pathogenicity of the 8 F. solani isolates Table 1 were tested first on cabbage seeds. Petri dishes (9 cm diameter) contained 20-15 ml of the cultured media (PDA) were inoculated with 5 mm of 6 days old of each. F. solani isolates taken from near the edge of each colony placed in the central of the petri dishes. 4 days later, cabbage seeds were superficially steriled with sodium Hiboklorate (1% free chlorine) for two minutes and placed as a circle around each petri dishes, an average of 10 seeds were placed in each petri dishes and three replicates/treatment (F. solani isolates) in a total number of 48 petri dishes except the control treatment without F. solani all petri dishes were incubated at 25 \pm 1 c ° for 4 days then the percentage of seed germination was calculated for each isolates compared with the control treatment (Bollkan and Butler, 1974).

Pathogenicity test of F. solani isolates on sour orange seeds

Sour orange seed from ripened non-infected fruits were extracted and cleaned thoroughly under running tap water until they get rid from adhesive substance and sterilized superficially with sodium Hiboklorat (1% free chlorine) for two minutes, then washed with distilled water and dry in the air. Sour orange seeds planted in plastic pots (15 cm diameter) containing sandy soil sterilized twice in autoclave (for 121 m, under pressure of 1.5kg/cm for one hour), 8 seeds were planted in each pot and three replicates / treatment (F. solani isolate) in total of 51 pot including 3 pots for control. A 7.5 gm. of each F. solani isolate culture on millet seeds were added to each pot and mixed with the sand soil, except for the control treatment (Kuthair, 2007). All pots was

transferred to a suitable place in a greenhouse and watered regularly with distilled water, and checked for seed germination. After the germination of all seedling in the control treatment, the numbers of seedling was calculated in each treatment and compared with the control treatment to calculate the percentage of seed germination. Complete random design (CRD) and statistical analysis program –SAS (2012) were used to compare the differences between treatments (Least significant differences (L.S.D. P. 0.05).

Results and Discussion

Isolation and Identification of F. solani from sour orange roots

Results of the isolation of *F. solani* from sample of sour orange roots which showed symptoms of typical symptoms of dry root rot and brown coloration of infected roots is present in all roots samples collected from the orchard regions in Baghdad province, with a high appearance rates between (93.3-100) % (Table 1) *F. solani* isolates taken from root orchards of Jadriyah (JA1), Tarmiyah (TM1) and Hamdaniya (HA1) have reached 100%. Microscopic examination showed that *F. Solani* have formed the three usual types of spores: Microconidia, Macroconidia and Chlamedospores. Using the taxonomic key of Booth s (1977) show it belong to *F. solani*.

It is clear from these results and the appearance of the fungus, *F. solani* at high rates, it might be the dominant fungus in the roots of the sour orange that show the typical symptom of citrus root rot, especially citrus orchards in Iraq suffered from stress factors such as drought and lack of regular irrigation during hot summer months, which may help in the development of the disease (Kuthair, 2007; Olsen, *et al.*, 2000; O'Connell, 2006). These findings are consistent with previously results found by many Iraqi researchers who found *F. solani* in high rates in citrus roots samples collected from different governorates of Iraq (Abbas, *et al.*, 1990; Al-Hakim, 1975; Mohammad, *et al.*, 1981; Al-Heeti, *et al.*, 1995; Kuthair, 2007).

Pathogenicity test of F. solani isolates on cabbage seeds

Result in Table (1) show a significant difference on the effects of *F. solani* isolates that collected from the different Sour orange orchids around Baghdad province on the germination of the cabbage seeds. Where percentage

of seed germination ranged between (3.3 - 30) % Compared with 96.6% for the control treatment.

Result indicated a significant differences in effects of F. solani isolates on the seed germination taken from the same regions of Taji and Tarmiyah, where Taji isolates amounting to 30 and 10% for TG1 and TG2 respectively, while it was 23.3 and 6.66% for Tarmiyah isolates TM1 and TM2. There were no significant differences was observed between the rests of the isolates obtained from the same area of the other 6 regions within the province of Baghdad. All F. solani has caused a significant reduction in the percentage of seed germination where the mean germination rates ranged from (4.95 - 24.95) %, Compared with 96.64%, in the control treatment. The lowest percentage of seed germination of 4.95 % recorded for Jadriyah (JA) isolates (Fig. 1), followed by isolates RS, TU, DO, TM which amounted to percentage of germination in 8.33, 13.3, 13.3 and 14.98 % respectively, and there in no significant differences between these isolates on their effect on seed germination. It was the highest percentage of germination of 20, 21.6, and 24.95 % for isolates TG, AC and HA respectively. Based on these Pathogenicity test, The 8 F. solani isolates can be divided into two groups, The first group which include isolates JA, RS, TU, DO and TM which germination rates ranged between (4.95 – 14.98) % on average of 10% , The second group includes isolates TG , AC and HA in which the proportions of germination high (20, 21.6 and 24.95) % reached twice what in the first group.

Table 1 Percentage of *F. solani* isolate on sour orange roots and their effect on Cabbage seed germination.

| Region | Isolates | % of F. solani appearance on sour orange roots | Percentage of cabbage seed germination | |
|---------------------|-------------|---|--|----------|
| | | | % seed germination / isolates | Mean |
| Al –Jadriyah (JA) | JA1 | 98.3 | 6.6 | |
| | JA2 | 100 | 3.3 | 4.95 |
| Al –Rashdiya (RS) | RS1 | 96.6 | 10 | |
| | RS2 | 98.3 | 6.6 | 8.33 |
| Al –Tuwaythah (TU) | TU1 | 95 | 10 | |
| | TU2 | 96.6 | 16.6 | 13.3 |
| Al-Dora (DO) | DO1 | 98.3 | 10 | |
| | DO2 | 95 | 16.6 | 13.3 |
| Al-Tarmiyah (TM) | TM1 | 100 | 23.3 | |
| • | TM2 | 96.6 | 6.66 | 14.98 |
| Al-Tajii (TG) | TG1 | 93.3 | 30 | |
| • , , | TG2 | 98.3 | 10 | 20.0 |
| Agriculture College | AC1 | 93.3 | 16.6 | |
| (AC) | AC2 | 95 | 26.6 | 21.6 |
| Al-Hamdaniya (HA) | HA1 | 100 | 26.6 | |
| • , , | HA2 | 96.6 | 23.3 | 24.95 |
| | Control | | 96.6 | 96.66 |
| | L.S.D. 0.05 | | 10.663 * | 10.479 * |

The effects on germination of cabbage seeds have been used by many researchers to evaluate the pathogenicity of different isolates of many fungal plant diseases (Bollkan and Butler, 1974; Hasson, 2005). In a similar study Kuthiar (2007) refers to the existence of significant differences between 4 F. *solani* isolates obtained from sour orange orchard collected from 4 provinces of between (4-53) % Several studies indicated that the different effects of *F. solani* isolates in the germination of cabbage and cauliflower seeds may be due to three reasons:

1-First is the genetic difference between fungus isolates of different regions (Nelson, *et al.*, 1983; Brasilerio, *et al.*, 2004; Juber and Hasson, 2006; Kuthair, 2007; O Connell;, 2006).

2-Producing a toxic metabolite which inhibit seed germination (Juber and Kuthair,2009; Jain and Thapliyal,1980), and finally the ability of theses isolates to produce analyst enzymes (cellulolytic enzyme, protease), Mycotoxins (Fusarubin and Javanicin) and other toxins (Baker *et al.*,1981;Nemec,1995; Nelson and Hansen,1997). The cell wall leading to

rotting and decomposition of seeds and lack of germination (Bateman and Lumsden, 1965; Pannecoucque and Hofte, 2009; Kuthair, 2007).

Pathogenicity test of F. solani isolates on sour orange seeds

Results in table (2) indicated a significant reduction in the percentage of the sour orange seeds germination grown in contaminated pots with 8 different F. solani isolates from different regions of Baghdad province, which ranged between (30 - 70)% compared with 96.66% in the control treatment. The lowest percentage of seed germination obtained for the two Jadriyah isolates amounted for 30, 46.6 % for JA1 and JA2 respectively and average mean of 38.3% which was significantly different from all other isolates obtained from other regions (Fig. 1). According to the results of the statistical analysis of the impacts of these isolates in the germination of the sour orange seeds we can divided these isolates into 3 main groups, the first group include Al-Jadriyah isolates JA1 and JA2 which considered the most effective and the high pathogenicity where germination did not exceed 38.3%, while the second group where germination rates where decreased to 50% which include isolates taken from Rashdiya region (RS), Tarmiyah (TM), Tuwaythah (TU) and the Collage of Agriculture (AC) which the germination rates between (48.3 – 55.1)%.

The third group which have the least effect where germination rate decreased (63.3-66.6)% for the isolates of Dora (DO), Taji (TG) and Hamdaniya (HA) respectively. There were many reasons about the effects of F. solani on the germination of the sour orange seeds, the most important is the ability to produce analyst enzymes, (Cellulotic enzyme, Protease), Mycotoxins (Fusarubin, Javaicin) and other toxins (Baker, et al., 1981; Obaid, 2004; Nemec, et al., 1991).

Based on these two pathogenicity test, Al-Jadriyah (JA) isolates have been chosen to conduct the next subsequent experimentation due to their high effect (Pathogenicity) on germination of the cabbage and sour orange seeds.

Table 2 Pathogenicity test of *F. solani* isolates on sour orange seeds.

| | Percentage of sour orang seed germination/ | | | |
|-------------------|--|---------|--|--|
| Region / isolates | isolate | Mean | | |
| JA1 | 30 | | | |
| JA2 | 46.6 | 38.30 | | |
| RS1 | 46.6 | | | |
| RS2 | 50 | 48.30 | | |
| TM1 | 43.3 | | | |
| TM2 | 60 | 51.65 | | |
| TU1 | 56.6 | | | |
| TU2 | 50 | 53.30 | | |
| AC1 | 70 | | | |
| AC2 | 43.3 | 55.15 | | |
| DO1 | 60 | | | |
| DO2 | 66.6 | 63.30 | | |
| TG1 | 60 | | | |
| TG2 | 70 | 65.55 | | |
| HA1 | 66.6 | | | |
| HA2 | 66.6 | 66.60 | | |
| Control | 96.6 | 96.66 | | |
| L.S.D. 0.05 | 7.925 * | 6.478 * | | |

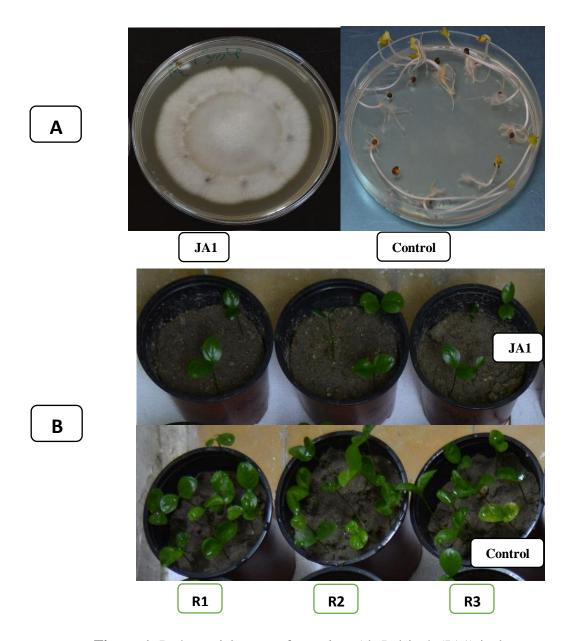


Figure 1 Pathogenicity test of F. solani Al- Jadriyah (JA1) isolate on Germination of: (A) Cabbage seeds, (B) Soure orange seeds

Acknowledgement

The authors would like to offer particular thanks to Dr. Feras and Dr. Salam and their collogues responsible for plant pathology Lab., Plant Protection Research Station, Ministry of Agriculture for their help and cooperation in completing this research.

References

- Abbas, A. H., F. A. Abdul-Fatah and M.N. Moheei. (1990). The role *of Fusarium solani* in slow die back of citrus, Iraqi journal of life sciences, 10:27-40.
- Al-Hakim, A.M.H. (1975). Occurrence rootstocks reaction, and control of the Citrus Nematode. *Tylenechulus semipenetrans* Cobb. In Iraq. M.Sc. thesis, Plant Port. Dept. Univ. of Baghdad, Iraq. 71PP.
- Al-Heeti, A. A., N. Mahdi, B. Hamid and W. Mohsson. (1995). Autumn blight, their Causes, nature of occurrence and control, The Iraqi journal of Agriculture Science's, 26:162-174.
- Baker, R. A., J. H. Tatum and S. Nemec. (1981). Toxin production by *Fusarium solani* from fibrous root of blight diseased citrus. Phytopathology. 71: 951-954.
- Bateman D. F. And R. D. Lumsden. (1965). Relation of calcium content and nature of Pectin substances in bean hypocotyls of different ages of *Rhizoctonia solani*, Phytopathology, 55:734-738.
- Bollkan, H.H and E.E Butler. (1974). Studies on Heterokaryosis Virulence of *Rhizoctonia solani*. Phytopathology., 64: 513-522.
- Booth, C. (1977). *Fusarium*. Laboratory guide to the identification of the major Species. Commonwealth Mycological Institute, Kew, Survey, England, 58 pp.
- Brasileiro , B.T.R.V. ; M.R.M.Coimbra ; M.A.M.Jr and N.T.oliveria. (2004). Genetic variability within *Fusarium solani* specie's as revealed by PCR-Finger-printing based on PCR markers. Brazilian Journal of Microbiology. , 35:205-210.
- Domsch , K. H. , W. Games and T. Anderson. (1980). Compendium of soil fungi . Academic Press , London , pp. 858.
- Farr, D. C., G.F. Bills, G.P. Chamuris , A.Y. Rossman. (1989). Fungi on Plants and plant products in the United States. APS press. ST. PAUL. MIN. 1251 PP.
- Hasson , I.K . (2005). Biological and chemical control to the potato stem , Ph. D. thesis , College of Agriculture, Univ. of Baghdad.
- Jain, R. K. and P. N. Thapliyal. (1980). Toxic metabolites from *Rhizoctonia solani* Kuhn: production and possible role in pathogenesis, Indian journal Expt. Biol., 18:316-318.
- Juber, K.S. and I. K. Hasson. (2006). Evaluation the efficiency of some Inducing compound chemicals and biocontrol agents in Controlling black scurf disease of potato under greenhouse and field conditions, The Iraqi Journal of Agricultural Sciences.

- Juber, K. S. and W. M. Kuthair. (2009). The relation between *Fusarium solani* casual of citrus root rot toxins and its pathogenicity, The journal of Agricultural Sciences, 40, (3): 63-77.
- Kamel , M. , M.N. Shatta and M.Z. Shanawanai. (1973). Histopathological studies on the hypocotyls of lentils infected by *Fusarium solani*. Z. Pflanzenkr. Pflanzenschutz 80: 547-550.
- Khosrow C., B. Salleh and L. Zakaria. (2015). Morphological and Phylogenetic Analysis of *Fusarium solani* Species Complex in Malaysia, Microb. Ecol., 69:457–471.
- Kumar, S., Thind T. S. and Chander M. (1995). Morphogenic and pathogenic variations in *Gloeosporium ampelogphagum*. Indian Phytopathology. 48: 331-334.
- Kunta; B. Salas; M. Gonzales and JV da Graca. (2015). First report of citrus dry root rot caused by *Fusarium solani* on sour orange rootstock in Texas, Journal of Citrus Pathology. J Cit Pathol. Iocv_journalcitruspathology_27974.
- Kuthair, W.M. (2007). Integrated control for citrus, root rot disease caused by *Fusarium solani*. Ph.D. Dissertations College of Agriculture, Univ. of Baghdad.119.pp.
- Lozovaya, V.V.; A.V.Lygin; O.V.Zernova; S. Li; J.M. Widholm and G.L. Hartman. (2006). Lignin degradation by *Fusarium solani* f.s. *glycines*. Plant Dis., 9:77-82.
- Mohammad, H.Y., S.I. Husain and A.J. Al-Zarai. (1981). Nematode found and Fungi flora associated with Citrus aurantium L. at seven different orchards of Nineveh and Diyala provinces of Iraq. Mesopotamia Journal of Agriculture 16: 185-205.
- Nalim F.A, Samuels, GJ, Wijesundera R.L, Geiser D.M. (2011). New species From the *Fusarium solani* species complex derived from perithecia and soil in the Old World tropics. Mycologia, 103:1302–1330.
- Nelson, P. E. Toussoun, T. A., and Marasas.W. F. O. (1983). Fusarium species: An Illustrated Manual for Identification. Pennsylvania State University Press, University Park. 193 pp.
- Nelson, B.D. and J.M. Hansen. (1997). Reaction of soybean cultivars to isolates of *Fusarium solani* from the red river valley. Plant Dis. 81:664-668.
- Nemec , S. , S. Jabaji Hair and P. M. Charest. (1991). ELISA and immunological detection of Fusarium solani produced napthazariin toxin in citrus tree in Florida. Phytopathology 81: 1497-1503.
- Nemec, S. (1995). Stress related compound in Xylem fluid of blight diseased citrus containing *Fusarium solani* naphthazarin toxins and their effect on the host. Can. J. Microbial., 41:515-524.
- Obaid, Q.M. (2014). Effectiveness of some biological agents to resist *Rhizoctonia solani* Kühn the cauliflower stem canker pathogen. M.Sc. thesis. Dept., of plant protection, College of Agriculture, 102p.
- O Connell , N. (2006). Dry root rot in citrus. University of California Cooperative Extension Tulare County. 9pp.
- Olsen, M., M. Matheron, M. McClure and Z. Xiong. (2000). Disease of 29Citrus in Arizona. Plant disease publications cooperative Extension, University of Arizona. 15pp.

- Pannecoucque ,J. and M. Hofte. (2009). Interactions between cauliflower and *Rhizoconia* anastomosis groups with different levels of aggressiveness, BMC Plant Biology, 9, 95 pp.
- Ravi, M. C. and M. R. Kumar. (2012). Studies on cultural, morphological variability in isolates of *Fusarium solani* (Mart.) Sacc., incitant of dry root- rot of Citrus Current Biotica 6(2): 152-162,
- Salam , A. H. (2009). Determination of variance resource and mechanism of pathogenicity for some *Beauveria bassiana* isolates and evaluation their efficiency in control corn stem borer *Sesamia certica* under field condition , Ph. D. thesis .Dept. of Plant Protection, College of Agriculture, Univ. of Baghdad
- SAS. (2012). Statistical Analysis System, User's Guide. Statistical. Version 9.1th ed. SAS. Inst. Inc. Cary. N.C. USA.
- Sherbakoff , C.D. (1953). *Fusarium* associated with citrus feeder roots in Florida. Phytopathology 43: 395-397.
- Tatum, J. H, and R. a. Baker. (1983). Naphthoquinones produced by *Fusarium solani* Isolated from citrus, Phytochemistry, 22:543-547.

(20 July 2016; accepted 5 September 2016)