Antifungal activities of *Chaetomium brasilense* CB01 and *Chaetomium cupreum* CC03 against *Fusarium oxysporum* f.sp. *lycopersici* race 2

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The antagonistic fungi of *Chaetomium brasilense* CB01 and *Chaetomium cupreum* CC03 were proved to antagonize *F. oxysporum* f.sp. *lycopersici* NKSC02 race 2 caused tomato wilt of sida abd cherry varieties. The bioactivities test demonstrated the antagonistic activity of *Ch. brasilense* CB01 and *Ch. cupreum* CC03 to inhibit the conidial production of *F. oxysporum* f. sp. *lycopersici* race 2. To elucidate the control mechanism involved in the inhibition of *F. oxysporum* f. sp. *lycopersici*, crude extracts of *Ch. brasilense* CB01 and *Ch. cupreum* CC03, were confirmed for antifungal activity against of *F. oxysporum* f. sp. *lycopersici* race 2. The other control mechanism involved in releasing antibiotic substances to inhibit *F. oxysporum* f. sp. *lycopersici* race 2. All tested crude extracts of *Ch. brasilense* CB01 and *Ch. cupreum* CC03, were significantly inhibited conidia production of *F. oxysporum* f. sp. *lycopersici* race 2. It is indicated that crude extracts from hexane, EtOAc and MeOH from *Ch. brasilense* CB01 inhibited *F. oxysporum* f.sp. *lycopersici* race 2 at the ED₅₀ of 29.87, 38.99 and 2.99 μg/ml, respectively. Crude extracts from hexane, EtOAc and MeOH from *Ch. cupreum* CC03 inhibited *F. oxysporum* f.sp. *lycopersici* race 2 at the ED₅₀ of 2.33, 2.38 and 2.65 μg/ml, respectively.

Key words: Chaetomium brasilense, Chaetomium cupreum, F. oxysporum f.sp. lycopersici race 2, fungal metabolites

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Introduction

Researches on natural products for antimicrobial against plant pathogens have been reported from several recent works. There are many new species of promising antagonists that can be used to control Fusarium wilt of tomatoes. The biocontrol agents and their bioactive compounds extracted from different species of antagonistic fungi were reported to inhibit the growth of many plant pathogenic fungi, including Fusarium wilt of tomato (Kanokmedhakul et al., 2006 and 2003; Thongsri and Soytong, 2004; Srinon et al. 2004, Suwannapong and Soytong, 2002 and Sibounnavong et al. 2009ab). The bioactive compounds, Trichotoxin A50 extracted from Trichoderma harzianum PC01; and Chaetoglobosin C extracted from Chaetomium globosum. compounds have also been reported to elicit resistance or immunity in plants by inducing oxidative burst in plant cells (Nuchdonrong et al. 2004; Soytong et al. 2001). The metabolites from fungi become one of potent antifungal against several plant pathogens. Crude extracts of Trichoderma hamatum WS01 and Penicillium sp.WS01 were reported to inhibit Fusarium oxysporum f.sp. cucumerinum and F. oxysporum f.sp. lycopersici isolated from wilt of cucumber and tomato (Srinon et al. 2006). Crude extracts from P. chrysogenum could protect cotton plants against wilt disease (F. oxysporum f. sp. vasinfetum and Verticillium dahlidae) and increases yield under field condition. (Dong et al. 2005, 2003; Dong and Cohen, 2001; Saidkarimov and Cohen, 2003) and Colletotrichum gloeosporioides (Soytong et al. 2005), Phytophthora parasitica (Meepeung and Soytong, 2004) and De Cal (2004) studied P. oxalicum to inhibit F. oxysporum f.sp. lycopersici and Botrytis cinerea. In addition, Gliocladium virens produced gliotoxin (Lumsden et al. 1992) and its properties against wood attacking fungi; Postia placenta and Neolentinus lepideus and Trametes versicolor and Phlebia brevisspora (Terry et al., 1996). Chulalak and Soytong (2006) reported that the bioactive compound extracted from Chaetomium cochliodes and Ch. cupreum inhibited plant pathogenic fungi, Phytophthora palmivora (root rot of pomelo) and Fusarium oxysporum f. sp. lycopersici (tomato wilt). Soytong et al. (2001) reported that the bioactive compound from Ch. cupreum inhibited the spore production of F. oxysporum which the ED₅₀ was 113.43 μ g/ml and inhibited the spore production of P. palmivora which the ED₅₀ was 53.46 µg/ml. Moreover, the bioactive compounds revealed that Ch. cupreum could reduce the sporulation of P. palmivora which the ED₅₀ was 279.67 μg/ml. With this, the ED₅₀ of crude extracts from Ch. cochlides was 323.01 µg/ml to inhibited F. oxysporum and the ED₅₀ of crude hence and ethyl acetate from Ch. cochliodes inhibited F. oxysporum were 203.64 and 416.41 µg/ml, respectively. A mechanism of antibiosis can occur during interactions involving low-molecular-weight diffusible compounds or production of antibiotics by biological control agents (Benítez et al. 2004). With this, the effective biological control agents produce several types of antibiotics to play important role in disease control (Lewis et al. 1989; Handelsman and Stabb, 1996). Specific species of fungi can produce specific metabolite that either impede spore germination as fungistasis, or kill the cells as antibiosis (Benítez et al. 2004). T. harzianum PC01 reported to produce trichotoxin A50 that it would induce resistant to mamy crops like tomato and potato etc. (Suwan et al. 2000). Ch. globosum can produce Chaetoglobusin C (Soytong et al. 2001) and Ch. cupream can produce rotiorinol (Kanokmedhakul et al. 2006). Antibiotics chaetoglobosin C and rotiorinol were reported to inhibit several plant pathogen e.g. F. oxysporum f. sp. lycopersici, C. gloeosporides and Phytopthora spp. (Soytong et al. 2001). The objective was to evaluate the biological activities of antagonists against F. oxysporum f. sp. lycopesici race 2 caused tomato wilt of sida abd cherry varieties.

Materials and methods

Pathogen to be tested: Fusarium oxysporum f.sp. lycopersici NKSC02 race 2 which pathogenic causing wilt to tomato var. Sida and Cherry from previous reports were used.

Effective antagonists to be tested:- Chaetomium brasilense CB01 and Chaetomium cupreum CC03 offered from Assoc. Prof. Dr. Kasem Soytong, Faculty of Agricultural Technology, King Mongkut's Institute of Technology Ladkrabang, Ladkrabang, Bangkok, Thailand were used.

Crude extraction:- Crude extracts from each antagonistic fungus were done follwed the method of Kanokmedhakul et al. (2006) and Moosophon et al. (2009). The fungi were cultured in potato dextrose broth (PDB) at room temperature for 30 days. The dried fungal biomass of each antagonistic fungus was ground and sequentially extracted with hexane, ethyl acetate and methanol. The solvents were then evaporated in vacuo to yield crude hexane, crude ethyl acetate (EtOAc), and crude methanol (MeOH) extracts, respectively.

Bioassays:- Crude extracts were assayed for inhibition of the most virulent isolate of *F. oxysporum* f. sp. *lycopersici* NKSC02 race 2. The experiment was conducted by using a factorial experiment in Completely Randomized Design (CRD) with four replications. Factor A represented the different solvents: A1 = crude hexane, A2 = crude ethyl acetate and A3 = crude methanol. Factor B represented the different concentrations: B1 = 0 μg/ml (control), B2 = 50 μg/ml, B3 = 100 μg/ml, B4 = 500 μg/ml and B5 = 1,000 μg/ml. Each crude extract was dissolved in 2% dimethyl sulfoxide and added to PDA before autoclaving at 121 °C (15 psi) for 30 minutes. To perform the assay,

a sterilized 3-mm diameter cork borer was used to remove agar plugs from the actively growing edge of the pathogen culture. An agar plug was transferred to the center of 5 cm diameter Petri dishes of PDA containing crude extract at each concentration and incubated at room temperature until the pathogen on the control plates had grown over the plate. Data were collected regarding the number of conidia produced by the pathogen and used to calculate the percentage of conidia inhibition. The effective dose (ED₅₀) was calculated using Probit analysis. The experiment was repeated twice.

Results

Chaetomium brasilense CB01 and Chaetomium cupreum CC03 at different concentrations of 0, 10, 50, 100, 500, and 1,000 g/ml were tested for inhibition of *F. oxysporum* f. sp. *lycopersici* NKSC02 which obtained from previous experiment. Hexane crude extract from *Ch. brasilense* CB01 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in colony diameter of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 3.67, 3.19, 2.67, 2.37 and 1.94 cm, respectively when compared to the control (0 μg/ml) of 5 cm. EtOAc crude extract from *Ch. brasilense* CB01 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in colony diameter of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 3.05, 2.92, 2.64, 2.27 and 2.22 cm, respectively when compared to the control (0 μg/ml) of 5 cm. MeOH crude extract from *Ch. brasilense* CB01 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in colony diameter of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 3.67, 3.50, 2.97, 2.77 and 2.22 cm, respectively when compared to the control.

Hexane crude extract from *Ch.cupreum* CC03 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in colony diameter of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 5.00, 4.87, 4.47, 4.45 and 4.12 cm, respectively when compared to the control. EtOAc crude extract from *Ch. cupreum* CC03 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in colony diameter of *F. oxysporum* f. sp. *lycopersici* NKSC02 which were 5.00, 3.92, 3.67, 3.54 and 3.40 cm, respectively when compared to the control. MeOH crude extract from *Ch. cupreum* CC03 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in colony diameter of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 4.47, 4.12, 3.74, 3.54 and 3.25 cm, respectively when compared to the control (Table 1 and 2).

Table 1. Effect of crude extracts from antagonistic fungi on mycelia growth of *Fusarium oxysporum* f.sp. *lycopersici* NKSC02

| Crude extracts | Colony diameter (cm) of Fusarium oxysporum f.sp. lycopersici at each concentration (µg/ml)) | | | | | | |
|----------------|---|--------|--------|--------|--------|--------|--|
| | 0 | 10 | 50 | 100 | 500 | 1000 | |
| C. brasilense | | | | | | | |
| Hexane | $5a^1$ | 3.67b | 3.19c | 2.67f | 2.37g | 1.94h | |
| EtOAc | 5a | 3.05cd | 2.92de | 2.64f | 2.27g | 2.22g | |
| MeOH | 5a | 3.67b | 3.50b | 2.97de | 2.77ef | 2.22g | |
| C. cupreum | | | | | | _ | |
| Hexane | 5a | 5.00a | 4.87a | 4.47b | 4.45b | 4.12b | |
| EtOAc | 5a | 5.00a | 3.92d | 3.67ef | 3.54f | 3.40g | |
| MeOH | 5a | 4.47b | 4.12c | 3.74e | 3.54f | 3.25 h | |

Average of four replications. Means followed by the same letters in each antagonist were not significantly different by DMRT at P=0.01.

Table 2. Effect of crude extracts from antagonistic fungi for percentage of colony inhibition growth of *Fusarium oxysporum* f.sp. *lycopersici* NKSC02

| Crude extracts of | Colony inhibition of Fusarium oxysporum f.sp. lycopersici (%) | | | | | |
|-------------------|---|--------|----------|----------|---------|--|
| | 10 | 50 | 100 | 500 | 1000 | |
| C. brasilense | | | | | | |
| Hexane | $26.5g^{1}$ | 36.0f | 46.5c | 52.5b | 61.0a | |
| EtOAc | 39.0ef | 46.0cd | 47.0c | 54.5b | 55.5b | |
| MeOH | 26.5 g | 30.0 g | 40.5 def | 44.5 cde | 55.5b | |
| C. cupreum | _ | _ | | | | |
| Hexane | 0.0h | 2.5h | 10.5g | 11.0g | 17.5f | |
| EtOAc | 0.0h | 21.5 e | 26.5 cd | 29.0 bc | 32.0 ab | |
| MeOH | 10.5 g | 17.5 f | 25.0 d | 31.0 b | 35.0 a | |

¹Average of four replications. Means followed by the same letters in each antagonist were not significantly different by DMRT at P=0.01

Hexane crude extract from *Ch. brasilense* CB01 at the concentrations of 10, 50, 100, 500 and 1000 µg/ml gave significantly different in spore production of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 21.5 x10⁷, 15.93 x10⁷, 14.0 x10⁷ and 2.16 x 10⁷ spore/ml, respectively when compared to the control (0 µg/ml) of 35.78 x10⁷ spore/ml. EtOAc crude extract from *Ch. brasilense* CB01 at the concentrations of 10, 50, 100, 500 and 1000 µg/ml gave significantly different in spore production of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 26.71 x10⁷, 19.61 x10⁷, 11.48 x10⁷, 5.35 x10⁷ and 4.40 x10⁷ spore/ml, respectively when compared to the control (0 µg/ml) of 36.24 x10⁷ cm. MeOH crude extract from *Ch. brasilense* CB01 at the concentrations of 10, 50, 100, 500 and 1000 µg/ml gave significantly different in spore production of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 11.83 x10⁷,

9.84 x10⁷, 8.52 x10⁷, 4.28 x10⁷ and 1.07 x10⁷ spore/ml, respectively when compared to the control (0 μg/ml) of 35.72 x10⁷ spore/ml. Hexane crude extract from *Ch. cupreum* CC03 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in spore production of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 15.93 x10⁷, 8.64 x10⁷, 6.82 x10⁷, 5.94 x10⁷ and 3.18 x10⁷ spore/ml, respectively when compared to the control (0 μg/ml) of 39.50 x10⁷ spore/ml. EtOAc crude extract from *Ch. cupreum* CC03 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in spore production of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 14.43 x10⁷, 8.87 x10⁷, 7.68 x10⁷, 4.48 x10⁷ and 2.40 x10⁷ spore/ml, respectively when compared to the control. MeOH crude extract from *Ch. cupreum* CC03 at the concentrations of 10, 50, 100, 500 and 1000 μg/ml gave significantly different in spore production of *F. oxysporum* f. sp. *lycopersici* NKSC02which were 13.93 x10⁷, 8.43 x10⁷, 6.16 x10⁷, 2.86 x10⁷ and 1.07 x10⁷ spore/ml, respectively when compared to the control (Table 3).

Table 3. Effect of crude extracts from antagonistic fungi against conidia production of *Fusarium oxysporum* f.sp. *lycopersici* NKSC02

| Crude extracts of | Number of conidia(x10 ⁷) of <i>Fusarium oxysporum</i> f.sp. <i>lycopersici</i> at each concentration (µg/ml) | | | | | | |
|-------------------|--|--------|---------|--------|--------|--------|--|
| | 0 | 10 | 50 | 100 | 500 | 1000 | |
| C. brasilense | | | | | | | |
| Hexane | $35.78a^{1}$ | 21.51c | 15.93d | 14.50d | 4.10gh | 2.16hi | |
| EtOAc | 36.24a | 26.71b | 19.61c | 11.48e | 5.35g | 4.40gh | |
| MeOH | 35.72a | 11.83e | 9.84ef | 8.52f | 4.28gh | 1.07i | |
| C. cupreum | | | | | | | |
| Hexane | 39.50b | 15.93d | 8.64fg | 6.82hi | 5.94i | 3.18k | |
| EtOAc | 38.47c | 14.43e | 8.87f | 7.68gh | 4.48j | 2.40k | |
| MeOH | 41.00a | 13.93e | 8.43 fg | 6.16i | 2.86k | 1.07 1 | |

Average of four replications. Means followed by the same letters in each antagonist were not significantly different by DMRT at P=0.01.

It revealed that crude extract at $1000 \,\mu\text{g/ml}$ from MeOH of *Ch. brasilense* CB01 gave significantly better inhibited spore production of *F. oxysporum* f.sp. *lycopersici* as 96.98 % better than crude extracts from EtOAc and MeOH which were 55.50 %. Crude extract at $1000 \,\mu\text{g/ml}$ from MeOH of *Ch. cupreum* CC03gave significantly better inhibited spore production of *F. oxysporum* f.sp. *lycopersici* as 97.37 % better than crude extracts from hexane and EtOAc which were 93.75 and 91.92 %, respectively. It is indicated that crude extracts from hexane, EtOAc and MeOH from *Ch. brasilense* CB01 inhibited *Fusarium oxysporum* f.sp. *lycopersici* race 2 at the ED₅₀ of 29.87, 38.99 and 2.99 $\mu\text{g/ml}$, respectively (Table 4). Crude extracts from hexane, EtOAc and MeOH from

Ch. cupreum CC03 inhibited Fusarium oxysporum f.sp. lycopersici race 2 at the ED₅₀ of 2.33, 2.38 and 2.65 μ g/ml, respectively (Table 4).

Table 4. Effect of crude extracts from antagonistic fungi for percentage of conidia inhibition of *Fusarium oxysporum* f.sp. *lycopersici* NKSC02

| Crude extracts | Conidia inhibition of Fusarium oxysporum f.sp. lycopersici (%) | | | | | |
|----------------|--|---------|---------|---------|--------|------------------------|
| | 10 | 50 | 100 | 500 | 1000 | ED ₅₀ μg/ml |
| C. brasilense | | | | | | |
| Hexane | $39.83k^{1}$ | 55.44i | 59.47h | 88.53c | 93.95b | 29.87 |
| EtOAc | 33.171 | 45.88j | 68.29g | 85.22d | 87.85c | 38.99 |
| MeOH | 66.87g | 72.38f | 76.09e | 87.98c | 96.98a | 2.99 |
| C. cupreum | | | | | | |
| Hexane | 59.64j | 78.23g | 82.71de | 84.38de | 91.92b | 2.33 |
| EtOAc | 62.46i | 76.92g | 82.17ef | 88.34c | 93.75b | 2.38 |
| MeOH | 65.99h | 79.41fg | 85.49d | 93.13b | 97.37a | 2.65 |

^TAverage of four replications. Means followed by the same letters in each antagonist were not significantly different by DMRT at P=0.01.

Discussion

The antagonistic fungi Ch. brasilense CB01 and Ch. cupreum CC03, were proved to antagonize F. oxysporum f.sp. lycopersici NKSC02 race 2 causing wilt to tomato var. Sida and Cherry. The antagonism test demonstrated the antagonistic activity of Ch. brasilense CB01 and Ch. cupreum CC03, to inhibit the conidial production of F. oxysporum f. sp. lycopersici NKSC02 between 63 - 77 %. Similar result was in accordance with the study from Charoenpoen et al. (2010) reported that Chaetomium lucknowense CLT significantly inhibited the mycelia growth and conidial production of F. oxysporum f. sp. lycopersici as 88.89 and 92.54 %, respectively. Furthermore, Sibounnavong et al. (2009) reported crude extacrts of Emericella nidulans strongly inhibited colonial growth and sporulation of F. oxysporum f. sp. lycopersici. Crude extracts of Ch. Brasilense CB01 and Ch. cupreum CC03 were confirmed for antifungal activity against of F. oxysporum f. sp. lycopersici NKSC02 race 2. The other control mechanism of *Ch. brasilense* CB01 and *Ch.* cupreum CC03 involved in releasing antibiotic substances to inhibit F. oxysporum f. sp. lycopersici. All tested crude extracts of Ch. brasilense CB01 and Ch. cupreum CC03 were significantly inhibited conidia production of F. oxysporum f. sp. lycopersici. This result was similar to the report of Charoenpoen et al. (2010) who stated that crude hexane, crude ethyl acetate and crude methanol from Ch. lucknowense CLT inhibited F. oxysporum f. sp. lycopersici NKSC01 with the ED₅₀ of 188, 209 and 212 µg/ml while in this study, crude extracts from methanol, ethyl acetate and hexane Ch. brasilense

CB01 inhibited the conidial production of different isolate of F. oxysporum f. sp. lycopersici NKSC02 race 2 with the ED₅₀ of 29.87, 38.99 and 2.99 µg/ml, respectively and crude extracts from methanol, ethyl acetate and hexane Ch. cupreum CC03 inhibited the conidial production of different isolate of F. oxysporum f. sp. lycopersici NKSC02 race2 with the ED₅₀ of 2.33, 2.38 and 2.65 µg/ml. Similar results were also reported by Srinon et al. (2006) and Sibounnavong et al. (2009) who stated that crude hexane, ethyl acetate and methanol extracts from E. nidulans inhibited the colony and sporulation of F. oxysporum f. sp. lycopersici. Moreover, Soytong et al. (2005) reported that crude ethyl acetate extract of Ch. globosum CG at 1000 µg/ml inhibited conidia production of this pathogen. As a result, Sibounnavong et al. (2009) reported that methanol crude extract from E. nidulans gave the highest inhibition of F. oxysporum f. sp. lycopersici. It is explained that ethyl acetate crude extract from E. rugulosa might have different fungal mrtabolites from methanol crude extract of E. nidulans as reported by Moosophon et al. (2006).

It concluded that *Ch. cupreum* CC03 can be produced some metabolites to inhibit *F. oxysporum* f. sp. *lycopersici* race 2 which Kanokmedhakul *et al.* (2006) found antifungal azaphilones from *Ch. cupreum* CC3003 effectively inhibited some human pathogens. Moreover, in this study *Ch. brasilense* CB01 proved to produce antifungal metabolites against *F. oxysporum* f. sp. *lycopersici* race 2 cause tomato wilt which it is the same isolate of reported *Ch. brasilense* CB01 by Khumkomkhet *et al.* (2009) found four new depsidones, mollicellins K-N which exhibited antimalarial activity against Plasmodium falcipurum and mollicellin K exhibited antmycobacterial activity against *Mycobacterium tuberculosis* and antifungal activity against *Candida albicans* and some cancer cell lines. The result of research finding would extend for testing to cotrol tomato wilt in the fields and further study would convey to apply these bioactive compounds as microbial elicitors to induce plant immunity.

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