VORACITY AND PREY PREFERENCE OF PHILIPPINE POPULATION OF TOXORHYNCHITES SPLENDENS WIEDEMANN (DIPTERA: CULICIDAE) AMONG AEDES SPP (DIPTERA: CULICIDAE) AND CULEX QUINQUEFASCIATUS SAY (DIPTERA: CULICIDAE)

Justine Bennette H Millado¹ and Augusto C Sumalde²

¹Department of Pest Management, Visayas State University, Visca, Baybay City, Leyte; ²Institute of Weed Science, Entomology, and Plant Pathology, College of Agriculture and Food Science, UP Los Baños, College, Laguna, Philippines

Abstract. Mosquito-borne diseases are a major burden in the Philippines for centuries due to the lack of effective and sustainable vector control measures. Sightings of the elephant mosquito, Toxorhynchites splendens in the country were recorded since the 1940's. It has not been mentioned, however, in any control programs to date. The prey consumption by this larval predator for *Aedes aegypti* (Linnaeus, 1762) (Diptera: Culicidae), Ae. albopictus (Skuse, 1894) (Diptera: Culicidae), and *Culex quinquefasciatus* Say (1823) (Diptera: Culicidae) was, therefore, studied under a "with choice" and "without choice" situation to initially assess its capability under local conditions. The average cumulative voracity of larval *Tx. splendens* was higher for Ae. aegypti than for Ae. albopictus at all densities, with 70% consumed by the fourth instar. An increase in predation relative to increase in density was observed for all Toxorhynchites instars. Consumption of males and females did not vary in both prey species. Percent preys consumed per number of preys offered declined as density increased suggesting a satiation point. When offered a mix of different instars of Ae. aegypti larvae, Tx. splendens first and second instar larvae consumed the younger instars, but fourth instar *Tx. splendens* preferred older larvae and pupa, and when offered a mix of larvae belonging to three different species, the majority of *Tx. splendens* instars preferred *Ae. aegypti* and *Ae. albopictus* over *Cx. quinquefasciatus.* Our results suggest that *Tx. splendens* has very good potential in controlling dengue-carrying mosquitoes under Philippine conditions.

Keywords: *Aedes* spp, *Toxorhynchites splendens*, mosquito control, prey preference, voracity

INTRODUCTION

The Philippines is placed fourth highest from 2008-2012 on the list of countries

Correspondence: Justine Bennette H Millado, Department of Pest Management, Visayas State University, Visca, Baybay City, Leyte, 6521-A Philippines.

E-mail: justinebennette@gmail.com

with incidence of dengue and dengue hemorrhagic fever in Southeast Asia, with an average of 117,065 cases yearly (Edillo *et al*, 2015). More recently, Philippines' CDC reported 12 novel cases of the emerging Zika from various provinces (Santos, 2016). Since the establishment of a control program by the Department of Health, various entomological surveys of communities around the country have been initiated, especially during vector surges in the wet months of August to November (Shultz, 1993). Attempts in controlling dengue remain challenging as existing vaccines still need improvement and therapy is only symptomatic (WHO, 2017). Population growth, urbanization, requirement for surveillance improvement, and limited success of vector control measures are correlated with increasing disease incidences (Bravo *et al*, 2014).

Control strategies in the country focus on the most widespread mosquito vectors *Aedes aegypti* (Linnaeus) (Diptera: Culicidae) and *Ae. albopictus* (Skuse) (Diptera: Culicidae) (WHO, 2007). Approaches include fumigation, use of permethrintreated bed nets and introduction of *Mesocyclops* sp and *Gambusia* sp. However, attempts to fully control vector populations have remained a major burden.

Alternative control measures using biological agents have recently gained interest especially those aimed at eliminating pre-adult stages. The predatory elephant mosquito, Toxorhynchites spp (Diptera: Culicidae), has shown great potential as larval carnivores of immature mosquitoes for the following reasons: (i) all larval stages are carnivorous and prey upon larvae and pupae of other mosquito species (Padgett and Focks, 1981); (ii) carnivorous larval stage may last for three weeks or more (Rubio and Ayesta, 1984); (iii) larvae are able to withstand desiccation and starvation for long periods and are able to survive on non-prev diet (Dodge, 1964; Trpis, 1979; Steffan and Evenhuis, 1981); (iv) larvae nearing pupation exhibit a "killing without eating" behavior in certain species (Corbet and Griffiths, 1963; Taylor, 1989; Collins and Blackwell, 2000); (v) adults are highly mobile and non-hematophagous; (vi) adult females may live up to seven weeks and have a lifetime egg production (Focks *et al*, 1979); and (vii) females oviposit on small natural and artificial water containers where target prey breeds and can be used in tandem with chemical control (Collins and Blackwell, 2000).

Mass releases of this predatory mosquito through inoculation or augmentation can be used to introduce or boost naturally occurring populations in the environment without causing any detrimental effect (Collins and Blackwell, 2000). Biocontrol attempts using Toxo*rhynchites* spp have long been carried out with some success in many neighboring countries such as Malaysia (Nyamah *et al*, 2011) and Singapore (Chan, 1968), and in Japan (Toma and Miyagi, 1992). However, no attempt has been initiated in the Philippines. Hence, the voracity of *Tx. splendens* was determined with and without choice of size and species of target preys.

MATERIALS AND METHODS

Mosquitoes

Starting population of Tx. splendens was established from immature stages collected in College of Forestry and Natural Resources, University of the Philippines Los Banos (UPLB), Laguna from August 2013 to February 2016. Oviposition traps composed of 24-cm wide 8-cm deep black pans fixed on top of a 3-foot wood block were set around the study area for gravid females. Sampling was conducted by scooping eggs into a water-containing vessel, and larvae and pupae were transferred using a modified asepto-syringe fitted with a rubber extension especially for water-containing habitats. All samples were brought to the laboratory at the Institute of Weed Science, Entomology and Plant Pathology, College of Agriculture and Food Science, UPLB for rearing. Species confirmation was performed by extracting DNA from 2-3 legs of some adult specimens and processed using Jena Bioscience[®] Animal and Fungal kit (Jena, Germany). PCR was performed as previously described (Kumar *et al*, 2007) and sent to 1st Base (Selangor, Malaysia) for sequencing. Rearing procedures were adapted from Furumizo and Rudnick (1978) for *Tx. brevipalpis* using *Ae. aegypti* as prey.

Ae. aegypti and *Ae. albopictus* egg strips were initially requested from the Insect Pathology and Storage Pests Laboratories, UPLB. Eggs were soaked in dechlorinated tap water and hatched larvae were fed with brewer's yeast. Adult females were given blood meals from a restrained guinea pig every two days. Egg rafts of *Cx. quinquefasciatus* were collected in ovitraps infused with rice hay. Rearing was conducted in cages covered with a black cloth and blood meals were provided by a restrained chick or dove.

Voracity for Aedes spp larvae assay

Experiments were divided into voracity (without choice) and host preference (with choice). For without choice situation, a newly-hatched Toxorhynchites wriggler was given daily five Ae. aegypti or Ae. albopictus larvae of the same instar. After 24 hours, the number of eaten larvae was counted and replaced with a new set of five preys. This was conducted throughout the larval period. This protocol was also performed at prey densities of 10, 20, 40, and 60. Tx. splendens larvae alone served as negative control, with mortality counts used to correct the number of eaten larvae in both trials using Sun-Shepard's formula (Puntener, 1981). Average number of dead individuals in the negative control was subtracted from the total average of wrigglers killed or consumed in

the corresponding treatment of the same replicate. Two trials were conducted for each prey density and species with at least 15 replicates for each experiment.

For the with choice situation, experiments were divided into two parts. In one part, *Tx. splendens* larvae were exposed to a set of different *Ae. aegypti* larval instars, and in the other part, predator larvae was exposed to a set of preys composed of a mix of three different species. *Tx. splendens* in the second, third, and fourth instar stages were given a diet of 10 *Ae. aegypti* daily until it reaches the desired age. Prior to the experiment proper, *Tx. splendens* wrigglers were starved for 12 hours after hatching or molting. The same protocol was also used for the negative control.

Preference for prey instar and species assay

A single first instar predator was introduced into a container with 20 *Ae. aegypti* larvae (five individuals from each of the four larval instars and five pupae). Uneaten preys after 24 hours were counted and this procedure was repeated until the predator molted and reached pupation.

In a single container, each *Toxorhychites* larva was given five individuals per species of larval prey (*Ae. aegypti, Ae. albopictus, Cx. quinquefasciatus*) belonging to the same instar. Uneaten preys after 24 hours were removed, killed, processed, and mounted on glass slides for species determination. This was carried out until the larva molted into the next instar and later pupated.

Statistical analysis

Shapiro-Wilk test in SPSS version 23.0 (IBM, Armonk, NY) was employed to test for non-normal data distribution of both voracity and preference. Additional tests were conducted using non-parametric Mann-Whitney *U* test involving data on

sex of adult mosquitoes and Kruskall-Wallis for variables related to density and species.

RESULTS

Voracity

Predatory consumption of *Ae. aegypti* by all *Tx. splendens* instars was positively influenced by prey density in both *Ae. aegypti* (Kruskal-Wallis, Sig. <0.0001) and *Ae. albopictus* (Kruskal-Wallis, Sig. <0.001) (Table 1). The average total number of wrigglers eaten by each *Tx. splendens* during its entire larval stage was higher for *Ae. aegypti* compared to *Ae. albopictus* larvae on a daily amount of 5, 10, 20, 40, and 60 preys. However, with the early instars, consumption was higher for *Ae. aegypti* plummeted during the third and fourth instars.

The fourth instar was the most voracious of the larval stages at all densities of both preys. A single fourth instar *Tx*. splendens consumed preys ranging from a minimum of 47 to a maximum of 222 Ae. aegypti and 39 to 222 Ae. albopictus over the entire duration of its stadium. The number of individuals eaten by the fourth instar constituted 59-66% (Ae. aegypti) and 59-70% (Ae. albopictus) of the total number of prey consumed during the entire larval period. However, as observed that on a daily basis, voracity was lower when given 5 prevs per day and consumption was comparable with the daily rates of the other instars. This lower daily consumption, consequently, was compensated by the long duration of the fourth instar in this type of experiment.

Although the mean consumption of the *Tx. splendens* larva on a daily basis and in total were positive in relation to density, the percent prey consumed over the total

number of prey given was descending for all instars. In the fourth instar, the decrease started at prey density of 10. This could be the point where the larva has reached its satiation. Male and female *Tx. splendens*, as determined after emergence, did not vary in the mean number of consumed *Ae. aegypti* and *Ae. albopictus* for all larval instars (Fig 1).

Preference for prey instar

First instar *Tx*. *splendens* was able to consume 3.36 first instar Ae. aegypti but not any fourth instars or pupae. Second and third instar Tx. splendens were able to attack fourth instar and pupae of Ae. *aegypti*, but to a lesser extent. Fourth instar *Tx. splendens*, on the other hand, readily preved upon fourth instar and pupae of Ae. aegypti and attacked first to second instars to a lesser degree. Non-parametric test using Kruskal-Wallis indicated that the voracity of different Tx. splendens instars was not the same for the different instars of Ae. aegypti (Sig. <0.0001). As the age of larval Tx. splendens increased, it favored late instars of Ae. aegypti, and conversely, younger Tx. splendens larvae preferred to attack younger instars of Ae. aegypti (Fig 2).

Preference for prey species

First, second, and fourth instar Tx. *splendens* larvae consumed, in general, more of the two *Aedes* spp larvae compared to *Cx. quinquefasciatus*. However, the third instar *Tx. splendens* preferred otherwise (Fig 3). These differences could have accounted for the indifference found on the preference of *Tx. splendens* over a specific species of prey (Kruskal-Wallis, Sig. = 0.98).

When grouped by instar, the consumption for each species is significantly different among *Ae. aegypti* (Kruskal-Wallis, Sig. <0.0001), *Ae. albopictus* (Sig.

	0
e	4
pl	ò
[a]	•÷
<u> </u>	0
	~ ~

Voracity of the four different Toxorhynchites splendens instars for Aedes aegypti and Ae. albopictus at different prey densities.

Prey density	First	بد	Secon	рı	Thire	T	Four	th	Tota	1
(number of prey/ Toxorhynchites										
wriggler)	$Mean\pm SD$	Total	$Mean\pm SD$	Total	$Mean\pm SD$	Total	$Mean\pm SD$	Total	$Mean\pm SD$	Total
5c	3 ± 1^{a}	7	3 ± 1	×	$2 \pm <1$	18	$2 \pm < 1$	48	$2 \pm < 1$	80
	$3\pm1^{ m b}$	8	2 ± 1		$2 \pm <1$	12	$2\pm <1$	39	$2\pm <1$	67
10^{d}	5 ± 2	6	4 ± 1	6	4 ± 1	26	6 ± 1	60	5 ± 1	134
	7 ± 2	6	6 ± 2	6	4 ± 1	11	6 ± 1	65	6 ± 1	94
20 ^e	5 ± 1	6	4 ± 1	11	6 ± 1	35	9 ± 1	119	7 ± 1	175
	7 ± 2	16	4 ± 1	13	4 ± 1	20	7 ± 1	94	6 ± 1	144
40^{f}	9 ± 2	18	10 ± 2	20	10 ± 1	46	15 ± 2	175	13 ± 1	259
	11 ± 1	21	10 ± 1	25	12 ± 2	52	15 ± 1	178	12 ± 1	277
60 ^g	10 ± 3	13	13 ± 2	45	13 ± 3	55	21 ± 2	222	17 ± 2	334
	11 ± 3	17	14 ± 2	23	13 ± 3	56	21 ± 1	222	15 ± 1	317
^a Aedes aegypti. ^b Ae. a $n = 34.^{e}Ae.$ aegypti, i preys given were of	<i>dbopictus.</i> ^c Total $\eta = 31$; <i>Ae. albopi</i>	number $n = n$	r of set-ups usi 33. ^f Ae. <i>aegypt</i> ne same larval	ng $Ae. a$ i, n = 30 instar as	egypti, n = 29; <i>i</i> ; Ae. albopictus, s the Toxorhync	Ae. albop $n = 30.$ shites wr	ictus, n = 35. ^d . ³ Ae. aegypti, n [:] iggler.	Ае. aegy = 32; Ае	pti, n = 52; Ae. c albopictus, $n =$	ılbopictus, 30. Aedes

SOUTHEAST ASIAN J TROP MED PUBLIC HEALTH



Fig 1–Mean daily consumption of *Aedes aegypti* and *Ae. albopictus* larvae by male and female *Toxorhynchites splendens* during the four larval stages at all prey densities. *Aedes* preys given were of proportionate size or the same larval instar as the *Toxorhynchites* wrigglers (*Ae. aegypti, n*=174; *Ae. albopictus, n*=163).

= 0.025), and *Cx. quinquefasciatus* (Sigs. = 0.002). Second and fourth *Tx. splendens* instars consumed more prey, regardless of the species, compared to the first and third instars.

DISCUSSION

Our findings on the voracity of *Tx*.

splendens for Ae. aegupti and Ae. albop*ictus* are comparable to that obtained by Begum et al (1988) where the average number of prev eaten during the larval life span is 257 ± 21. Trpis (1973) in Kenva showed Tx. brevipalpis consume 154 - 358 Ae. aegypti during the course of its larval stage. On the other hand, a lower number is obtained for Ae. albopictus in comparison to the 389 prey larvae eaten by Tx. splendens (Toma and Miyagi, 1992). Difference in the consumption was only more evident in the last two stadia and were more uniform in the first and second stadia in both males and females (Rubio and Avesta 1984). Differences in the range of minimum numbers of preys consumed could be attributed

to the number of larvae offered and effects of ambient temperature. Although Corbet (1963) stated that fourth instar *Tx. brevipalpis condrati* is able to reduce feeding at low food availability, the experimental conditions in the current study involved replenishment of eaten prey, thereby, even at a low prey density, *Toxorhynchites* larvae still had a constant supply of prey *per se*.



Fig 2–Prey preference of first to fourth *Toxorhynchites splendens* instar when given a mix of first to fourth instars and pupae of *Aedes aegypti* as prey within 24 hours. *Aedes* preys given were of proportionate size or the same larval instar as the *Toxorhynchites* wrigglers (n = 36).



Fig 3–Prey species preference of the four different larval instars of *Toxorhynchites splendens* when given a mix of *Aedes aegypti, Ae. albopictus,* and *Culex quinquefasciatus*. Preys offered were of proportionate size or the same larval instar as the *Toxorhynchites* wrigglers (n = 42).

The large number of prevs consumed by the fourth instar compared to the younger instars could be attributed to its higher nutritional requirement, especially the energy needed during the non-feeding pupal period (Crans and Slaff, 1977). Trimble and Smith (1978) reported the minimum amount of prev consumed by the fourth instar of Tx. brevipalpis larva is 60-70% of its total consumption during the whole larval stage. This last instar is considered to have the most intense phagoperiod according to the rate of biosynthesis across many species of mosquitoes (Timmerman and Briegel, 1998).

Foraging theory suggests that predators learn to adjust consumption based on the energy content and size of the prey in light of its specific nutritional requirements. Schmidt et al (2012) explained that the previous nutritional experience of generalist predators, such as the wolf spider, pre-determines its choice of prey. It has

long been understood that the potential energy provided by the target prey is the major factor influencing predator foraging behavior (Whelan and Schmidt, 2007). However, more in depth studies revealed that it could be further broken down into the prey's impact on the predator's growth, development, distribution, and survival (Schmidt *et al*, 2012). As a result, most predators regulate food intake to maximize nutrition and prevent excessive consumption of prey with less nutritional value. Even after capture, predators still regulate ingestion to match their metabolic intake (Mayntz *et al*, 2005).

In the field, *Toxorhynchites* larva food source is limited by the rate at which potential prev hatch and develops. These are highly influenced by the prey's oviposition rate as well as water level (Russo, 1986). The low rate of consumption exhibited by *Tx. splendens* in the study by Russo (1986) compared to other species, such as Tx. theobaldi (Dyar and Knab, 1906), could be attributed to the habitat where this species thrives. Unlike Tx. theobaldi has adapted to a drier environments, and therefore, consume large amounts of food during the short time available, while *Tx*. splendens has a constant supply of prey food in the wetter tropical countries.

The preference for a prey of the same instar or body proportion was also observed in *Tx. brevipalpis* Theobald (1901), where in the presence of younger instars, fourth instar *Toxorhynchites* preferred to consume third and fourth instar prey larvae and pupae (Lounibos, 1979). Also, the consumption of fourth instar *Ae. aegypti* is higher than of pupae and dramatically higher than of first instars when offered altogether (Padgett and Focks, 1981). However, when offered separately, *Tx. rutilus* is able to eat 93 first instars compared to 10 fourth instars and 7 pupae (Coquillet, 1896).

Toxorhynchites larvae do not actively seek out prev and prev capture mostly relies on the collisions between itself and its prev (Zuharah et al. 2015). Among the four larval instars and pupae offered, the fourth instar Aedes larva has the highest activity and often "grazes" on the body of *Toxorhynchites* larva, thereby increasing interaction compared to the more lethargic pupa (Zuharah et al, 2015; Nyamah et al, 2011). The preference over the younger prey instars was theorized to be based on the optimal foraging strategy, which explains that attacking a larger prey requires a lower attack rate. The decrease in time needed to assault a prev outweighs the added energy needed for a longer handling time. In this way, Toxorhunchites larva may have evolved this mode of predation as it is more economical as regards time and energy than stalking smaller preys even though they are easier to consume (Lounibos, 1979).

The voracity of *Tx. splendens* for *Ae*. aegypti and Ae. albopictus increased as it reached the second instar, then dropped during the third instar and slightly increased again as it reached the last instar. Predatory insects such as *Toxorhynchites* spp are expected to increase in voracity as they prepare for the high energyconsuming pupal stage as observed in most cases (Chan, 1968; Furumizo and Rudnick, 1978), making the low consumption during the third instar, a peculiarity. During this stage, larvae were observed to passively rest near the container's edge. This thigmotactic behavior is theorized to be a manifestation of anxiety (Schnörr et al, 2012). Also, unlike other instars that readily attack approaching preys, this instar prefers to observe passing Aedes larvae by moving its head towards the prey's direction without attacking. Although unclear, these behaviors could be attributed to the development of the compound eyes, which is usually initiated during this stage as observed with other dipterans (Wolff and Ready 1993; Ashburner *et al*, 2005). While *Toxorhynchites* larvae depend on movement-based signals (Zuharah *et al*, 2015) for detecting prey, this development of eyes may be a novel process in the growing larva, which could also explain its thigmotaxis.

Voracity of *Tx. splendens* for *Clue* was similar to that observed for the *Aedes* varied in such *Toxorhynchites* sp is able to consume 570 *Cx. quinquefasciatus* during its lifetime (Urmila *et al*, 2000), a number greater than most studies using *Aedes* (Steffan and Evenhuis, 1981; Toma and Miyagi, 1992). Comparing the three prey species in our study, *Cx. quinquefasciatus* was observed to be consumed the most, probably owing to its higher chance of contact stemming from its higher movement activity especially by the third instar larva.

In conclusion, predatory preference of *Tx. splendens* is an important factor which contribute to high reduction in Aedine populations. The use of this predatory mosquito will be instrumental to the success of any dengue and dengue hemorrhagic fever control effort in the Philippines.

ACKNOWLEDGEMENTS

The authors thank the DOST-ASTHRDP program, Dr BL Caoili and Dr PA Javier for providing the initial batch of *Aedes* eggs and technical assistance in the molecular work and Dr OK Bautista and all other readers for the improvement of the manuscript.

REFERENCES

- Ashburner M, Golic KG, Hawley RS. Drosophila. A laboratory handbook. 2. New York: Cold Spring Harbor Laboratory Press, 2005.
- Begum EM, Saleem N, Rahman MM, et al. The biology of *Toxorhynchites splendens* (Wiedemann) (Diptera: Culicidae) and its potentiality as a biological control agent to other mosquitoes. *Bangladesh Med Res Counc Bull* 1988; 14: 15-20.
- Bravo L, Roque VG, Brett J, Dizon R, L'azou M. Epidemiology of dengue disease in the Philippines (2000-2011): a systematic literature review. *PLOS Negl Trop Dis* 2014; 8: e3027.
- Chan KJ. Observations on *Toxorhynchites splendens* (Wiedmann) (Diptera: Culicidae) in Singapore. *Mosq News* 1968; 28: 91-5.
- Collins LE, Blackwell A. The biology of *Toxo-rhynchites* mosquitoes and their potential as biocontrol agents. *Biocont News Info* 2000; 21: 105-16.
- Coquillett DW. New Culicidae from North America. *Can Entomol* 1896; 28: 43-4.
- Corbet PS. Observations on *Toxorhynchites brevi*palpis conradti Grünb. (Diptera: Culicidae) in Uganda. *Bull Entomol Res* 1963; 54: 9-17.
- Corbet PS, Griffiths A. Observations on the aquatic stages of two species of *Toxorhynchites* (Diptera: Culicidae) in Uganda. *Proc R Entomol Soc London (A)* 1963; 38: 125-35.
- Crans WJ, Slaff ME. Growth and behavior of colonized *Toxorhynchites rutilus septentrionalis. Mosq News* 1977; 37: 207-11.
- Dodge HR. Larval chaetotaxy and notes on the biology of *Toxorhynchites rutilus septentrionalis. Ann Entomol Soc Am* 1964; 57: 46-53.
- Dyar HG, Knab F. The species of mosquitoes in the genus Megarhinus. *Smithsonian Miscel laneous Collections* 1906; 48: 241-58.
- Edillo FE, Halasa YA, Largo FM, *et al.* Economic cost and burden of dengue in the Philippines. *Am J Trop Med Hyg* 2015; 92: 360-6.

- Focks DA, Seawright JA, Hall DW. Field survival, migration and ovipositional characteristics of laboratory-reared *Toxorhynchites rutilus rutilus* (Diptera: Culicidae). *J Med Ent* 1979; 16: 121-7.
- Furuzimo RT, Rudnick A. Laboratory studies of *Toxorhynchites splendens* (Diptera: Culicidae): biological observations. *Ann Ent Soc Am* 1978; 71: 670-3.
- Kumar Np, Rajavel AR, Natarajan R, Jambulingam P. DNA barcodes can distinguish species of Indian mosquitoes (Diptera: Culicidae). J Med Entomol 2007; 44: 1-7.
- Lounibos LP. Temporal and spatial distribution, growth and predatory behavior of *Toxorhynchites brevipalpis* on the Kenya Coast. *J Anim Ecol* 1979; 48: 213-36.
- Mayntz D, Raubenheimer D, Salomon M, Toft S, Simpson S. Nutrient-specific foraging in invertebrate predators. *Science* 2005; 307: 111-3.
- Nyamah MA, Sulaiman S, Omar B. Field observation on the efficacy of *Toxorhynchites splendens* (Wiedemann) as a biocontrol agent against *Aedes albopictus* (Skuse) larvae in a cemetery. *Trop Biomed* 2011; 28: 312-9.
- Padgett PD, Focks DA. Prey stage preference of the predator *Toxorhynchites rutilus rutilus* on *Aedes aegypti*. *Mosq News* 1981; 41: 67-70.
- Puntener W. Manual for field trials in plant protection. 2nd ed. Basle: Ciba-Geigy, 1981.
- Rubio Y, Ayesta C. Laboratory observations on the biology of *Toxorhynchites theobaldi*. *Mosq News* 1984; 44: 86-90.
- Russo R. Comparison of predatory behavior in five species of *Toxorhynchites* (Diptera: Culicidae). *Ann Entomol Soc Am* 1986; 79: 715-21.
- Santos TG. Zika infects pregnant woman in Cebu. *Philippine Daily Inquirer.* 2016 Sept 27. [Cited 2017 Jan 23]. Available from: <u>http://newsinfo.inquirer.net/819327/zikainfects-pregnant-woman-in-cebu</u>
- Schultz GW. Seasonal abundance of dengue vectors in Manila, Republic of the Philip-

pines. Southeast Asian J Trop Med Pub Health 1993; 24: 369-75.

- Schmidt JM, Sebastian P, Wilder SM, Rypstra AL. The nutritional content of prey affects the foraging of a generalist arthropod predator. *PLOS One* 2012; 7: e49223.
- Schnörr SJ, Steenbergen PJ, Richardson MK, Champagne DL. [Abstract]. Measuring thigmotaxis in larval zebrafish. *Behav Brain Res* 2012; 228: 367-74.
- Steffan WA, Evenhuis NL. Biology of *Toxorhynchites. Ann Rev Entomol* 1981; 26: 159-81.
- Taylor DS. Preliminary field observations on the killing behavior of *Toxorhynchites amboinensis* larvae. J Am Mosq Control Assoc 1989; 5: 444-5.
- Timmerman SE, Briegel H. Molting and metamorphosis in mosquito larvae: a morphometric analysis. *Mitteilungen der Schweizerischen Entomologischen Gesellschaft* 1998; 71: 373-87.
- Toma T, Miyagi I. Laboratory evaluation of *Toxorhynchites splendens* (Diptera: Culicidae) for predation of *Aedes albopictus* mosquito larvae. *Med Vet Entomol* 1992; 6: 281-9.
- Trpis M. Interaction between the predator *Toxorhynchites brevipalpis* and its prey *Aedes aegypti. Bull World Health Organ* 1973; 49: 359-65.
- Trpis M. Development of the predatory larvae of *Toxorhynchites brevipalpis* on non-prey diet. *J Med Entomol* 1979; 16: 26-28.
- Trimble RM, Smith SM. Geographic variation in development time and predation in treehole mosquito, *Toxorhychites rutilus septentrionalis*. *Can J Zool* 1978; 56:1256-65.
- Urmila J, Ganesh KN, Vijayan VA. Feeding capacity of *Toxorhynchites* sp. larvae under laboratory conditions. *Insect Environ* 2000; 6: 116-7.
- Whelan CJ, Schmidt KA. Food acquisition, processing, and digestion. In: Stephen DW, Brown JS, Ydenberg RC, eds. Foraging: behavior and ecology. Chicago: University

of Chicago Press, 2007: 141-72.

- Wolff T, Ready DF. Pattern formation in the *Drosophila* retina. In: Bate M, Martinez Arias A, eds. The development of *Drosophila melanogaster*. Vol 2. New York: Cold Spring Harbor Laboratory Press, 1993; 1277-325.
- World Health Organization (WHO). Scientific Working Group Report on Dengue: meeting report, 1-5 October 2006. Geneva: WHO, 2007. [Cited 2012 Apr 23]. Available from: http://www.who.int/tdr/publica-

tions/publications/swgdengue2.htm

- World Health Organization (WHO). Dengue vaccine research. Geneva: WHO, 2017. [Cited 2017 Feb 05]. Available from: <u>www.</u> who.int/immunization/research/development/dengue_vaccines/en/
- Zuharah WF, Fadzly N, Yusof NA, Dieng H. Risky behaviors: effects of *Toxorhynchites splendens* (Diptera: Culicidae) predator on the behavior of three mosquito species. J *Insect Sci* 2015; 15:128.